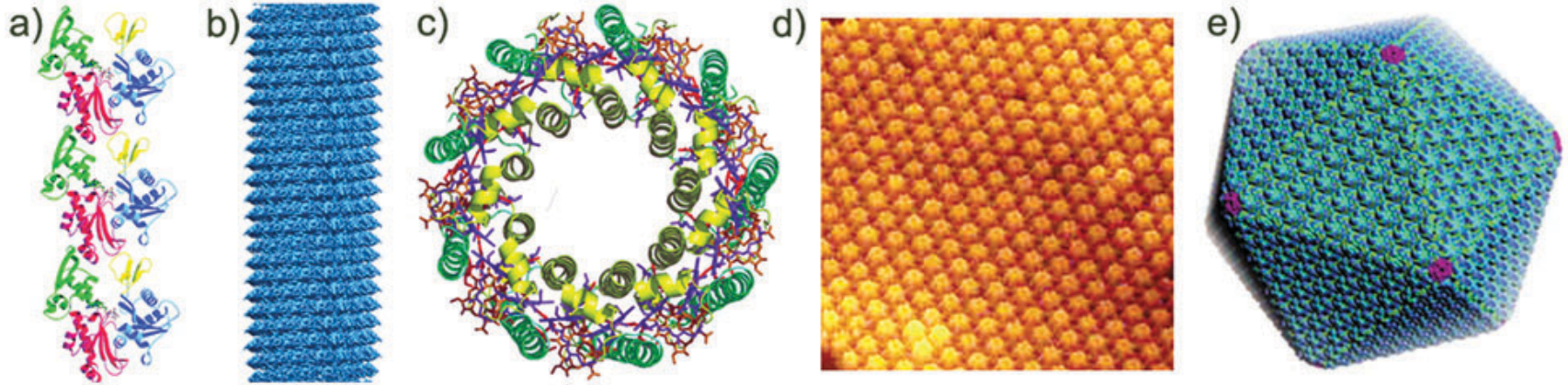


# Self assembly



# Shape in eukaryotic cells

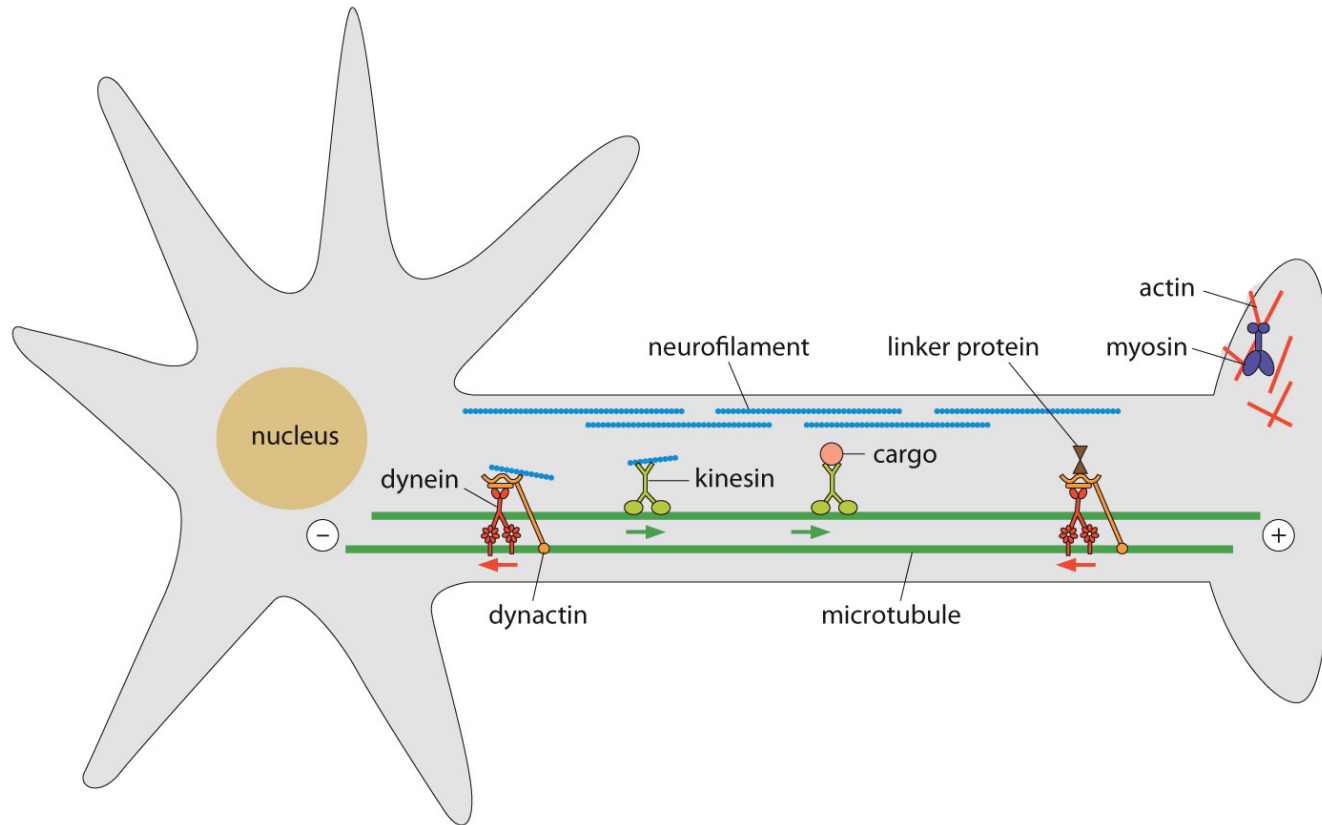


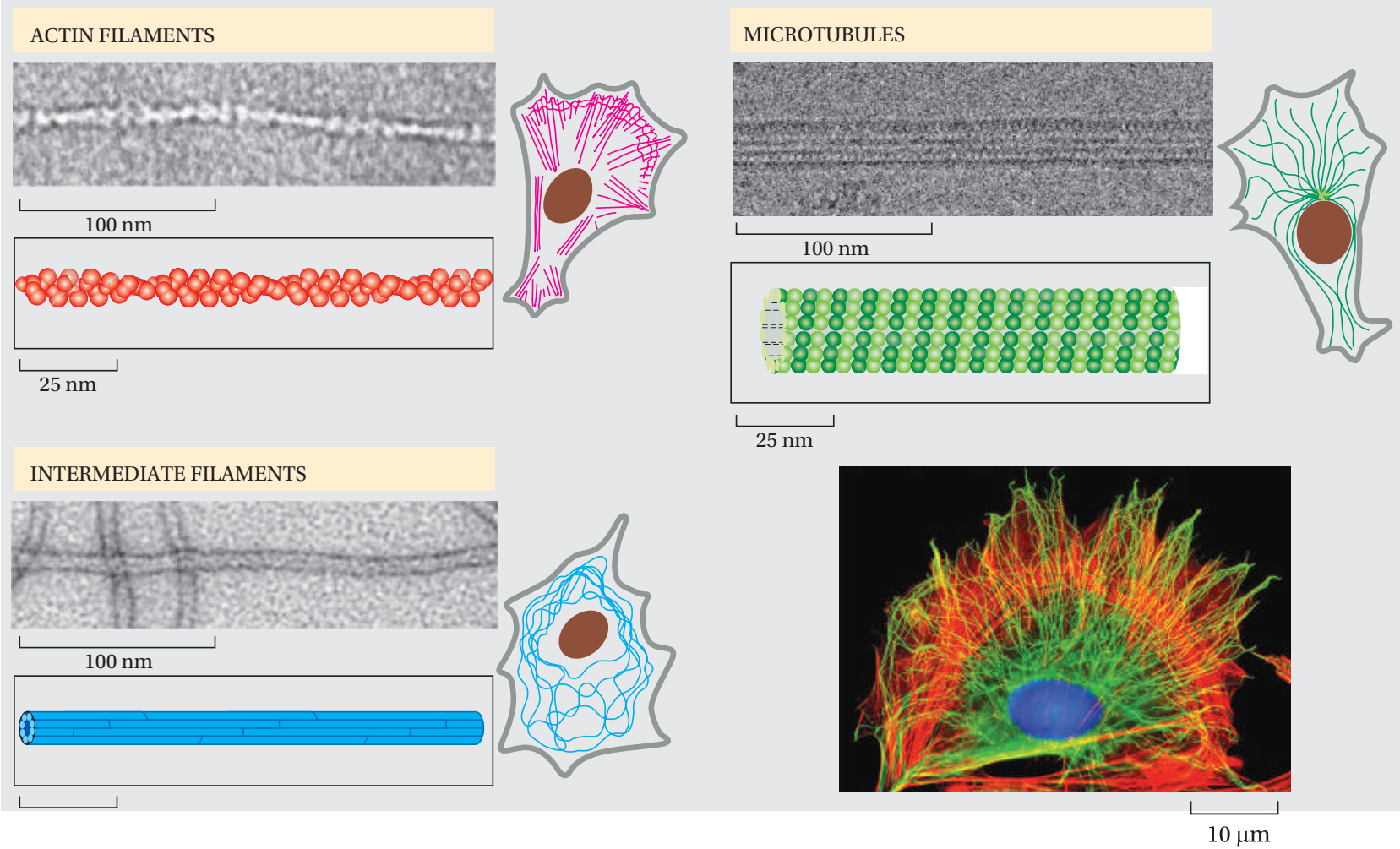
Figure 14.1 Molecular Biology of Assemblies and Machines (© Garland Science 2016)

Shape in eukaryotic cells is provided by the cytoskeleton that consists of actin, tubulin, and intermediate filaments.

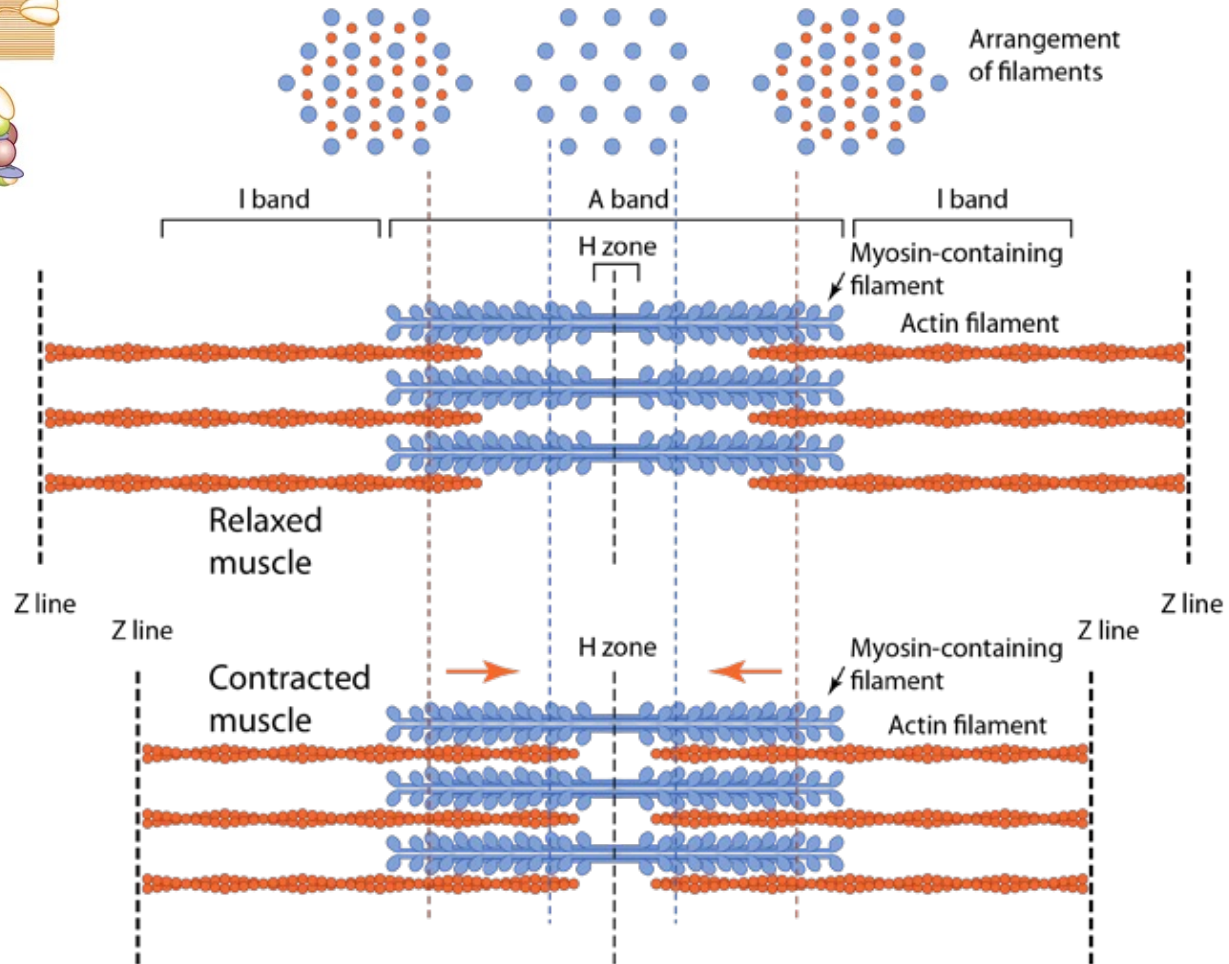
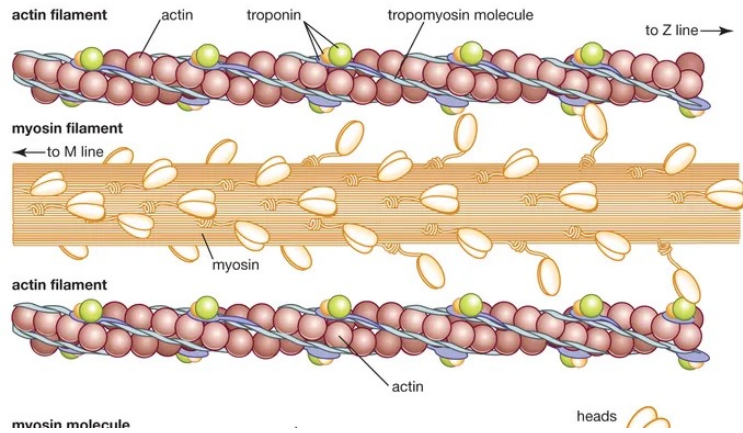
Bacterial cells come in a variety of different shapes, including spheres, rods, spirals, and crescents.

Shape is important for bacterial cells because it plays a role in cell division, helps to maximize the uptake of nutrients, and aids cell movement.

# Filaments and microtubules within the cell



# Actin





# Actin filament

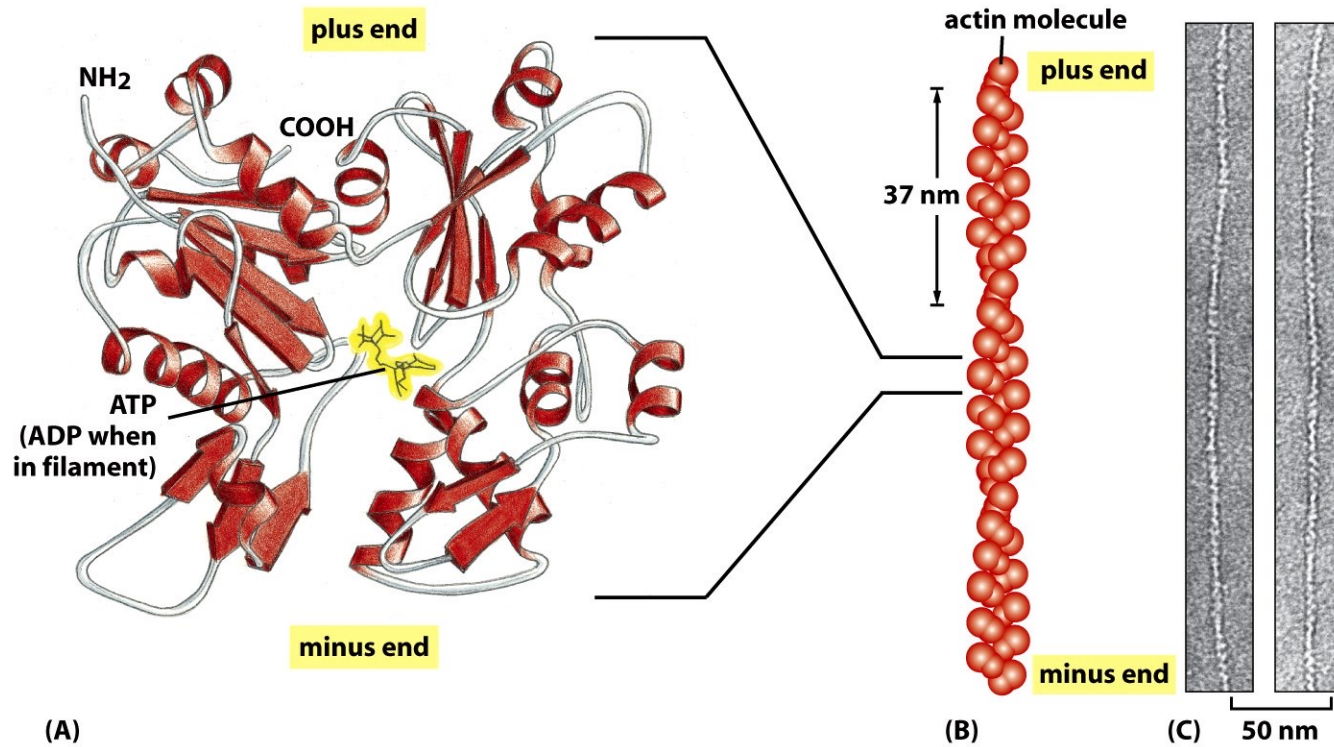
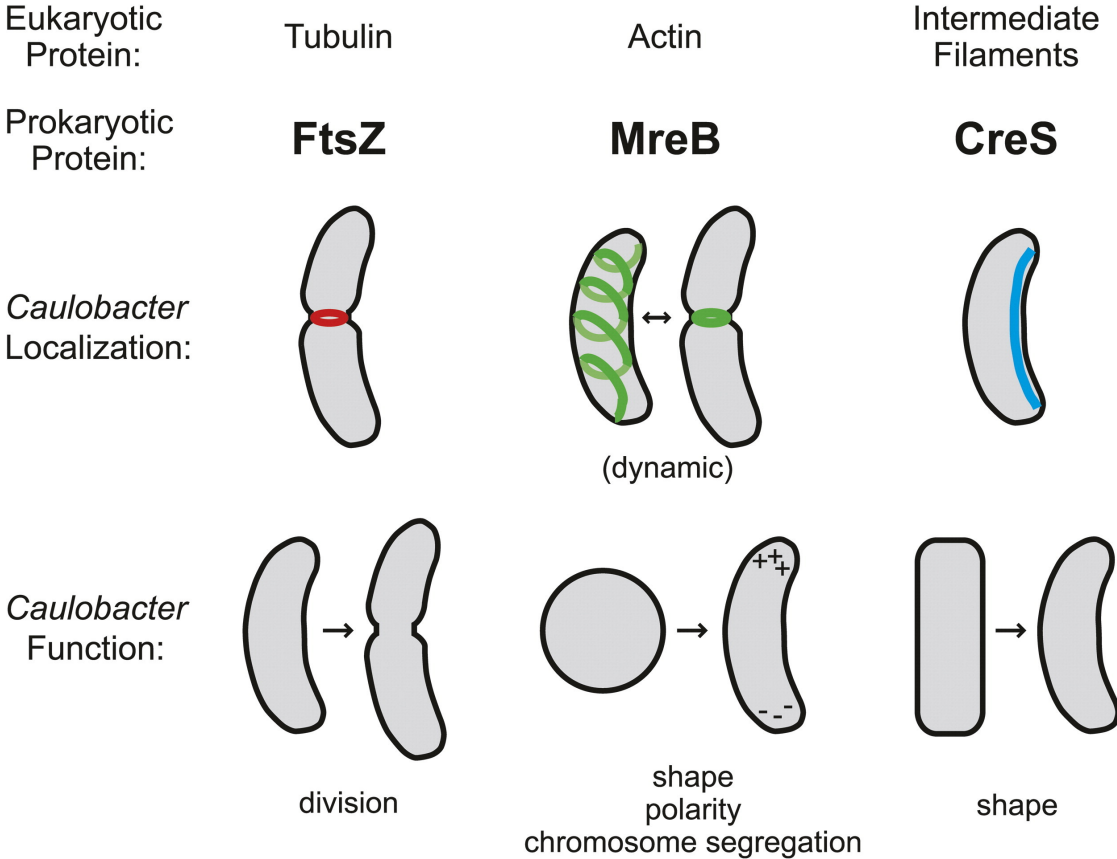


Figure 16-12 *Molecular Biology of the Cell* (© Garland Science 2008)

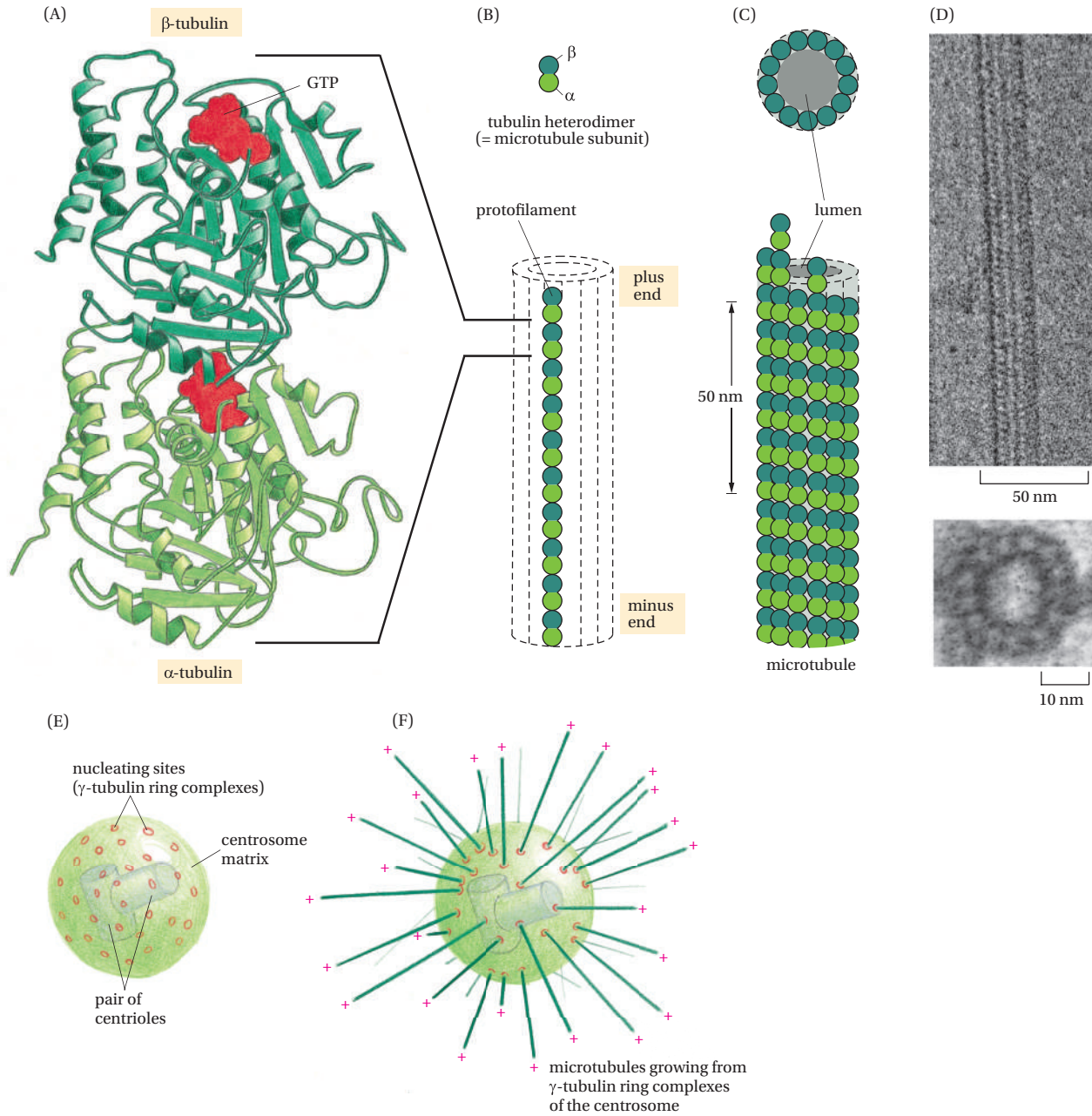
# Procaryotic actin

Prokaryotes also have a dynamic, filamentous network of proteins, which are homologous to the eukaryotic cytoskeletal elements.

In non-spherical bacteria, the actin homologue MreB is essential for shape maintenance as depletion of MreB through genetic knockouts or MreB-targeted drug treatment results in misshapen cells that eventually lyse



# microtubules



# Dynamic instability

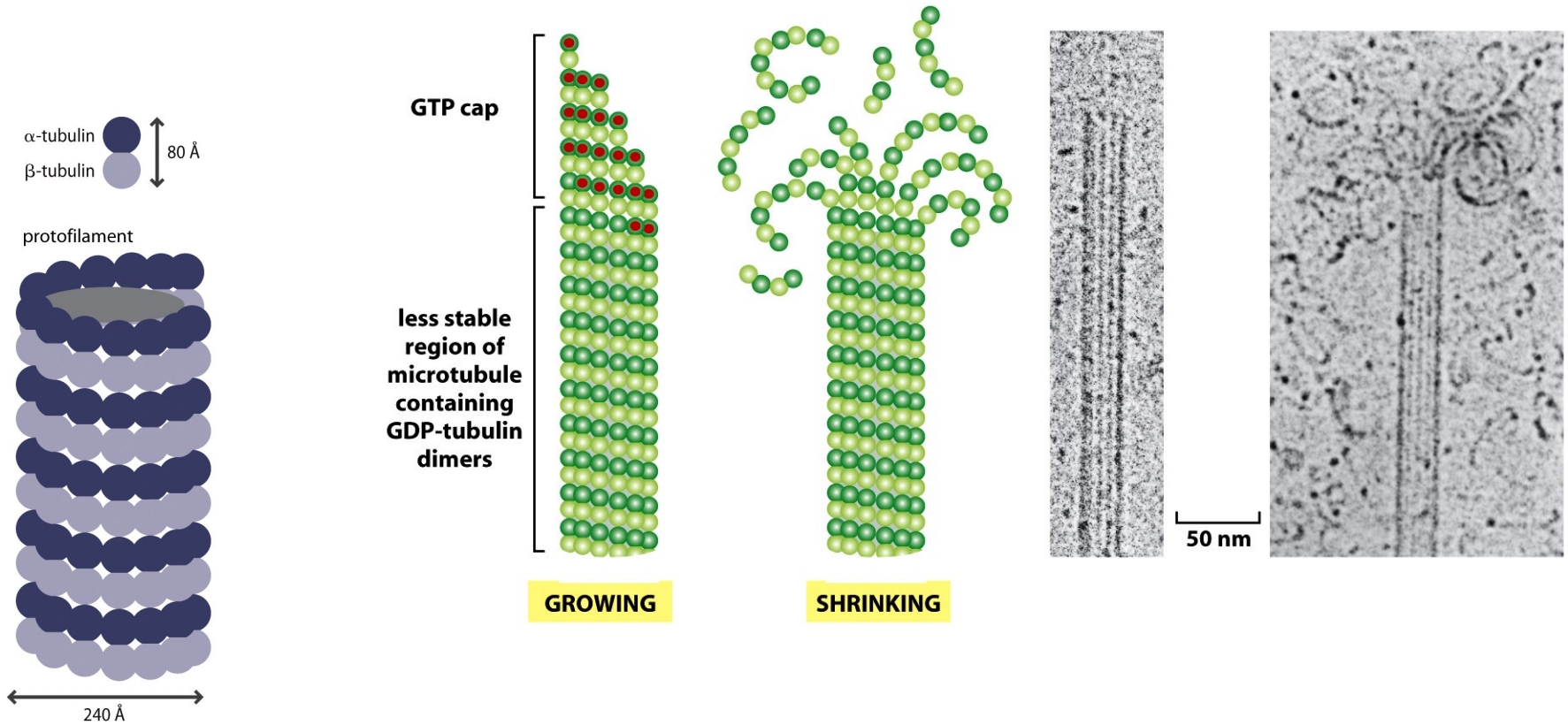


Figure 7.27 How Proteins Work (©2012 Garland Science)



# Tubulin interacting proteins

<b>TABLE 7.2 Proteins that interact with tubulin/microtubules</b>	
<b>Protein</b>	<b>Function</b>
$\gamma$ -TuRC	Initiates filament formation
MAP, XMAP215	Stabilizes filaments
Tau, MAP-2	Cross-links filaments in parallel rows
Stathmin, kinesin 13, katanin	Cuts or depolymerizes filaments
+TIP, plectin	Links filament to other proteins

Table 7.2 How Proteins Work (©2012 Garland Science)

# Fiber growth

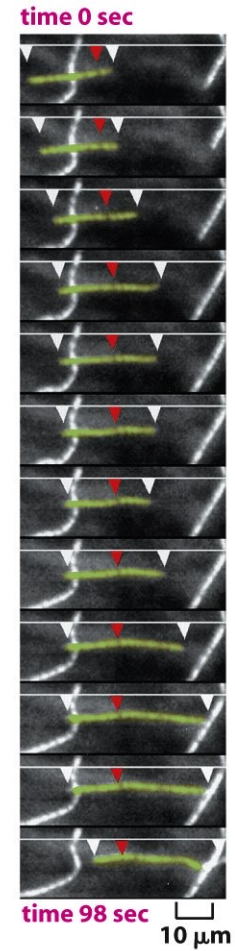
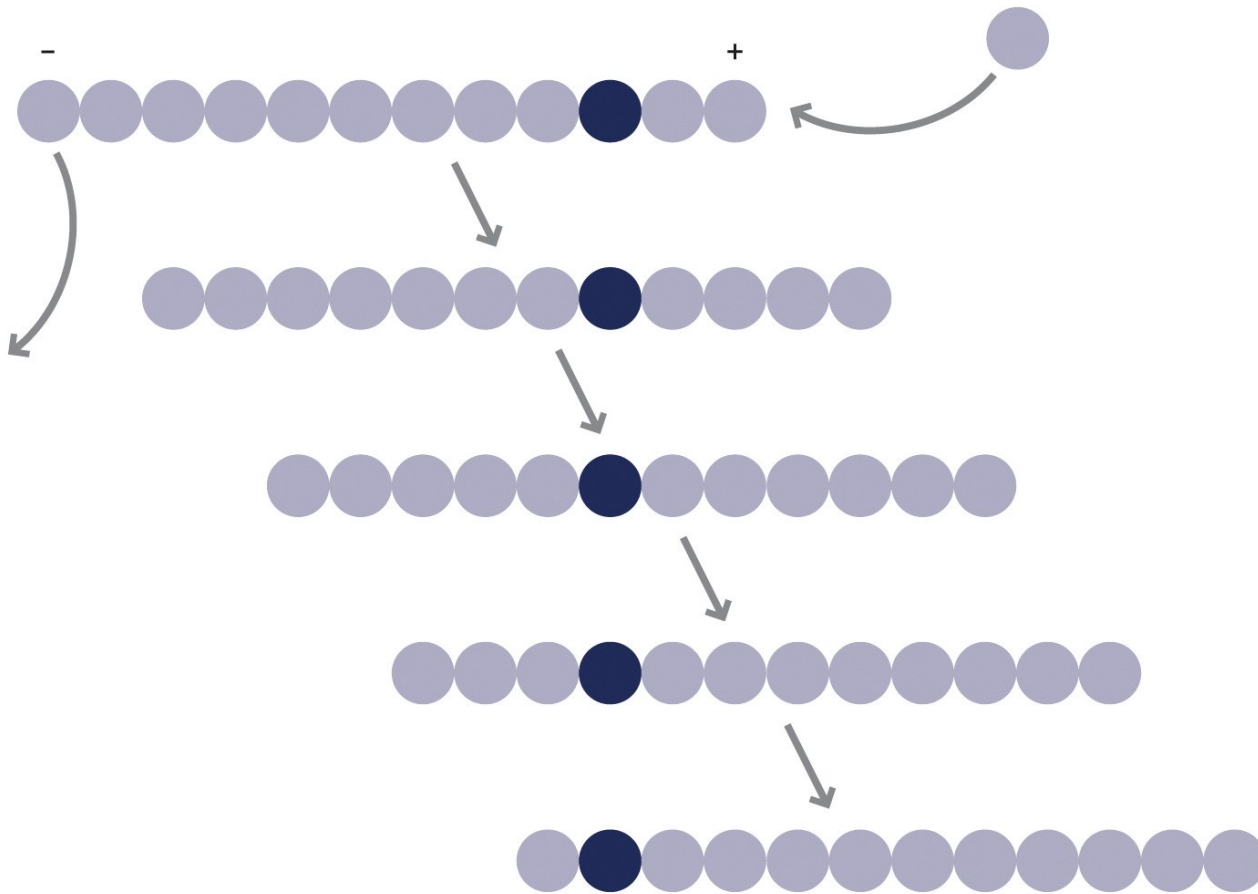
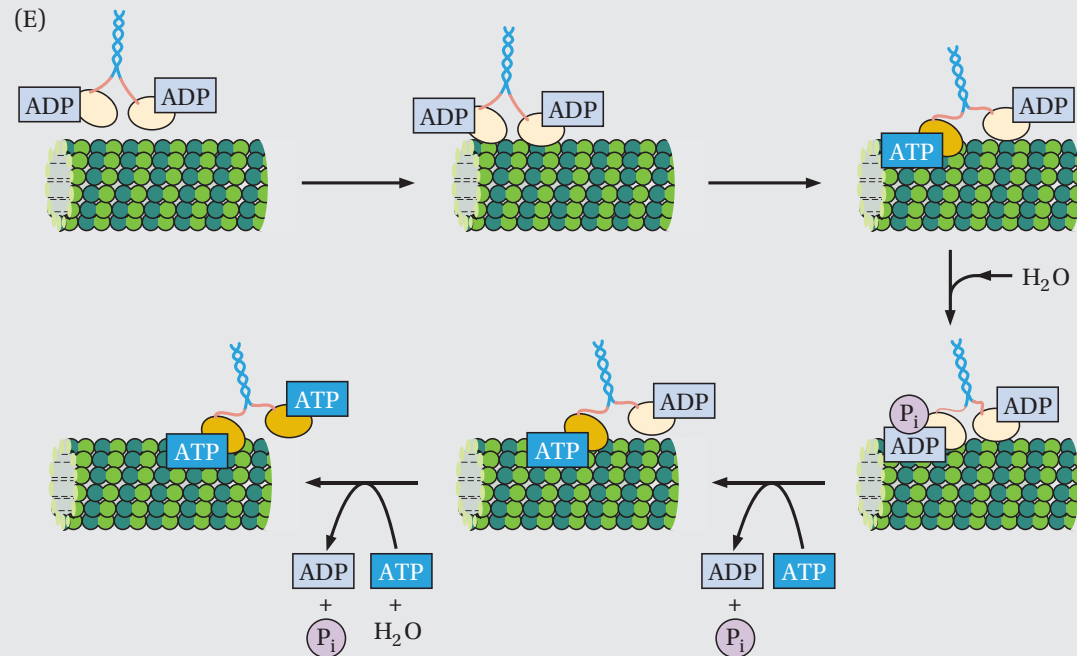
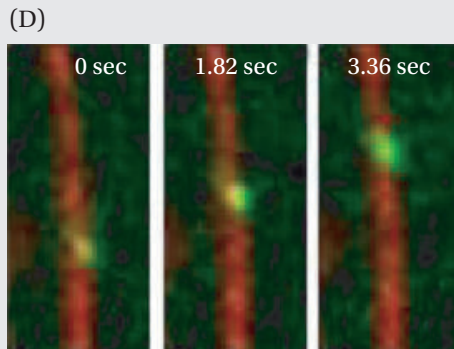
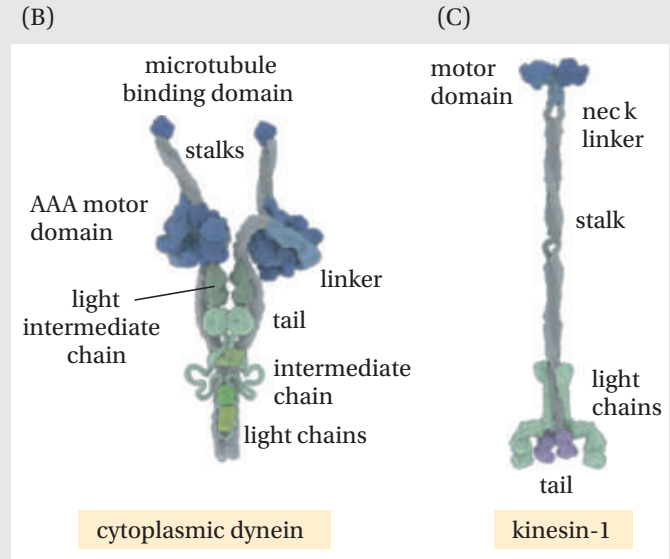
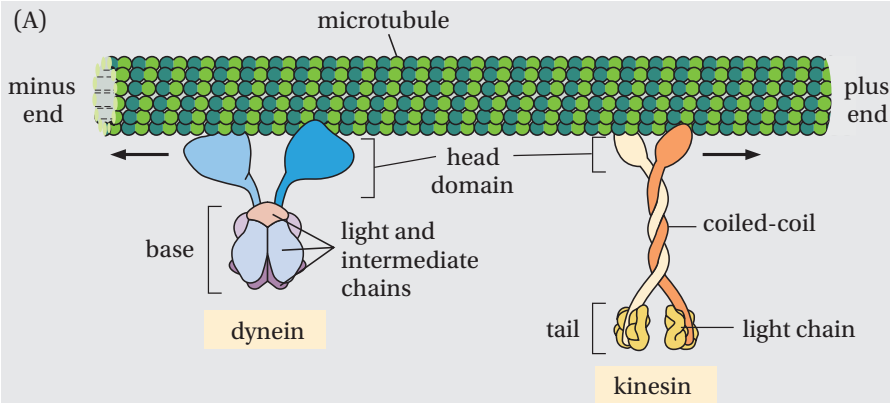


Figure 7.30 How Proteins Work (©2012 Garland Science)

# kinesins and dyneins



# Drugs

**Table 16–2 Drugs That Affect Actin Filaments and Microtubules**

<b>ACTIN-SPECIFIC DRUGS</b>	
<b>Phalloidin</b>	<b>binds and stabilizes filaments</b>
<b>Cytochalasin</b>	<b>caps filament plus ends</b>
<b>Swinholide</b>	<b>severs filaments</b>
<b>Latrunculin</b>	<b>binds subunits and prevents their polymerization</b>
<b>MICROTUBULE-SPECIFIC DRUGS</b>	
<b>Taxol</b>	<b>binds and stabilizes microtubules</b>
<b>Colchicine, colcemid</b>	<b>binds subunits and prevents their polymerization</b>
<b>Vinblastine, vincristine</b>	<b>binds subunits and prevents their polymerization</b>
<b>Nocodazole</b>	<b>binds subunits and prevents their polymerization</b>



# Fluorescence microscopy

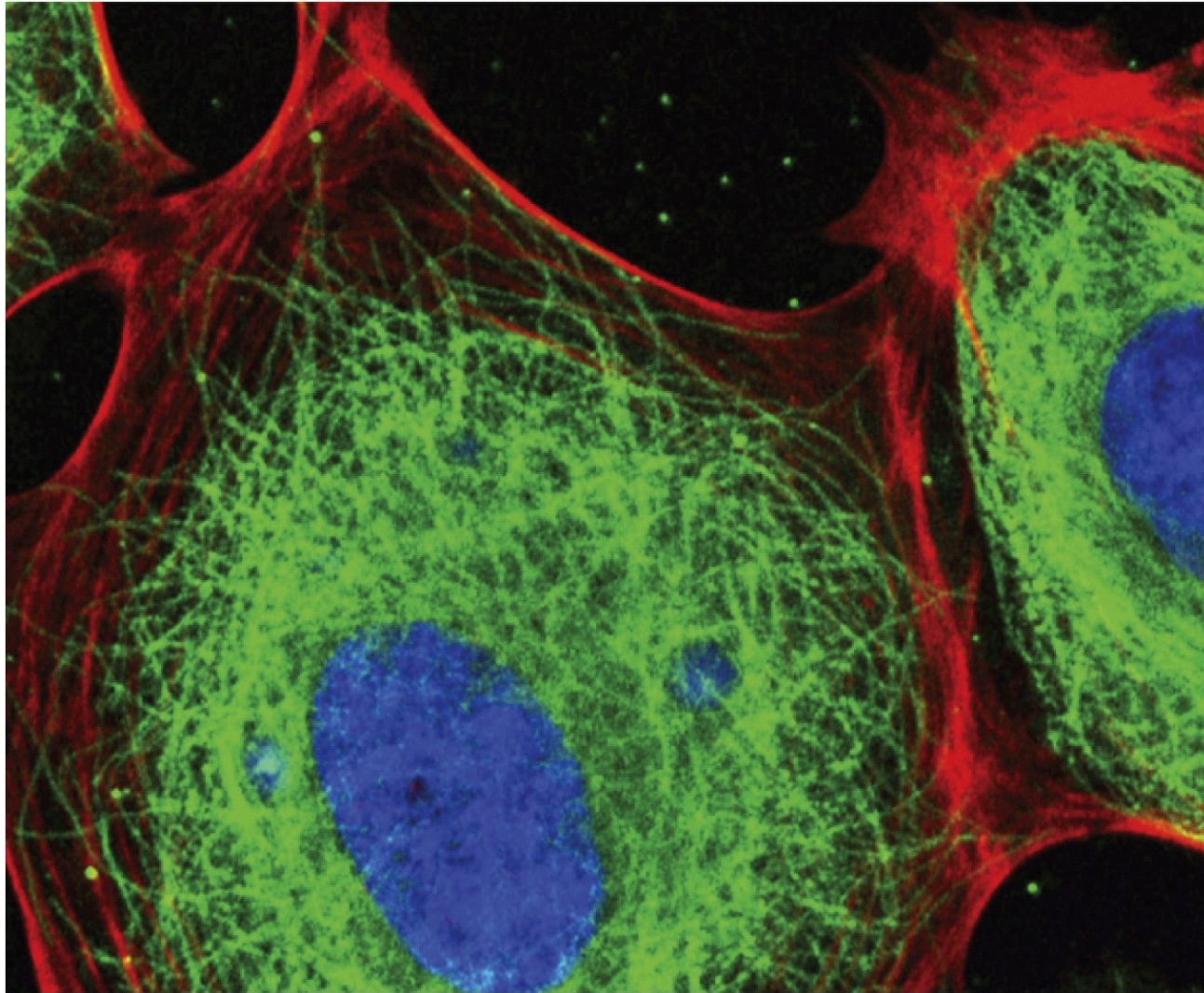
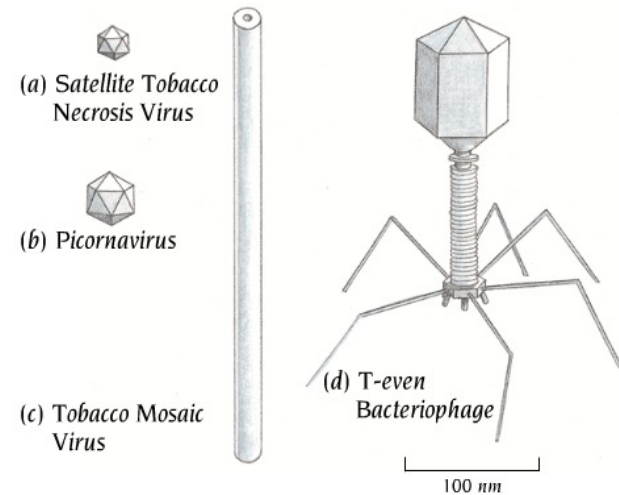
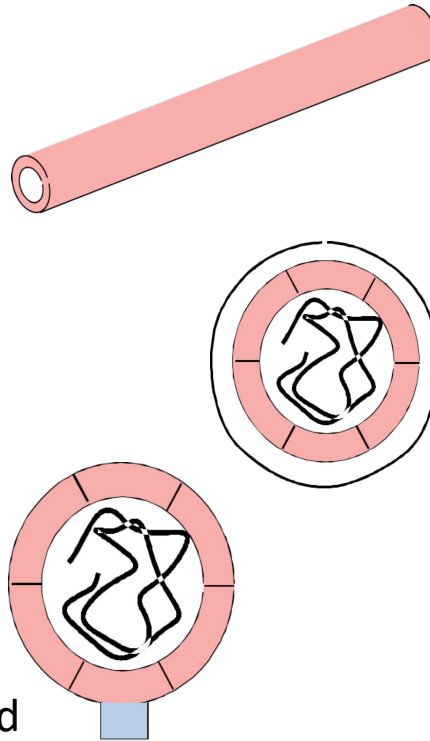


Figure 7.33 How Proteins Work (©2012 Garland Science)

# Virus Structure

# Virus Structure

- Size
  - 17 nm – 3000 nm diameter
- Basic shape
  - Rod-like
  - “Spherical”
- Protective Shell - Capsid
  - Made of many identical protein subunits
  - Symmetrically organized
  - 50% of weight
  - Enveloped or non-enveloped
- Genomic material
  - DNA or RNA
  - Single- or double-stranded



# Virus Structure

- Virus capsids function in:
  - Packaging and protecting nucleic acid
  - Host cell recognition
    - Protein on coat or envelope “feels” or “recognizes” host cell receptors
  - Genomic material delivery
    - Enveloped: cell fusion event
    - Non-enveloped: more complex strategies & specialized structures



# Viruses

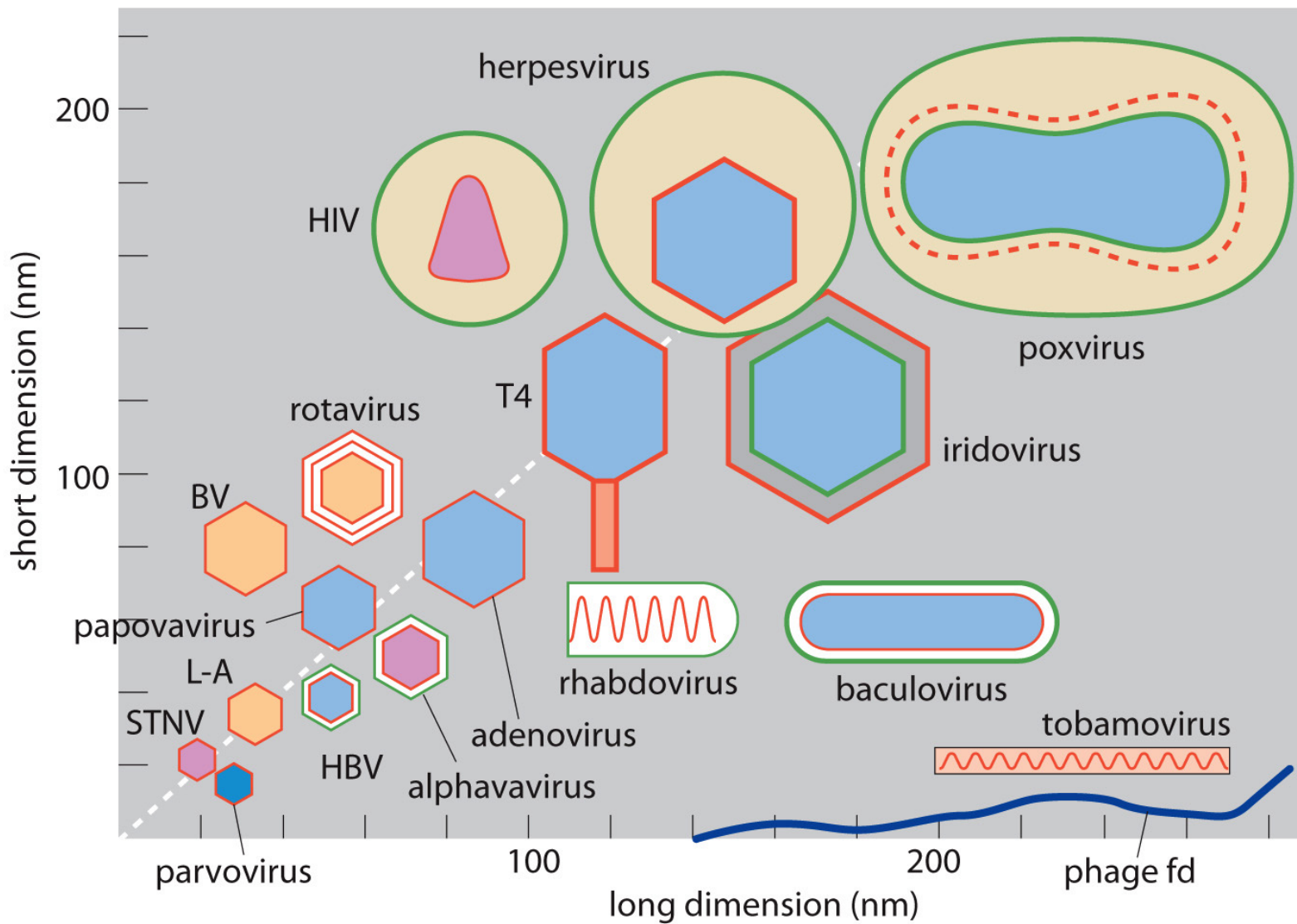
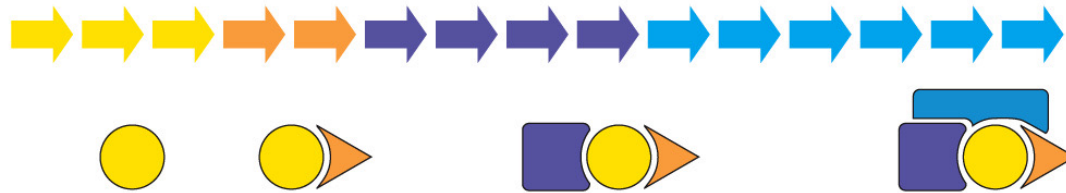


Figure 8.4 Molecular Biology of Assemblies and Machines (© Garland Science 2016)

# assembly pathways

linear assembly pathway



branched assembly pathway

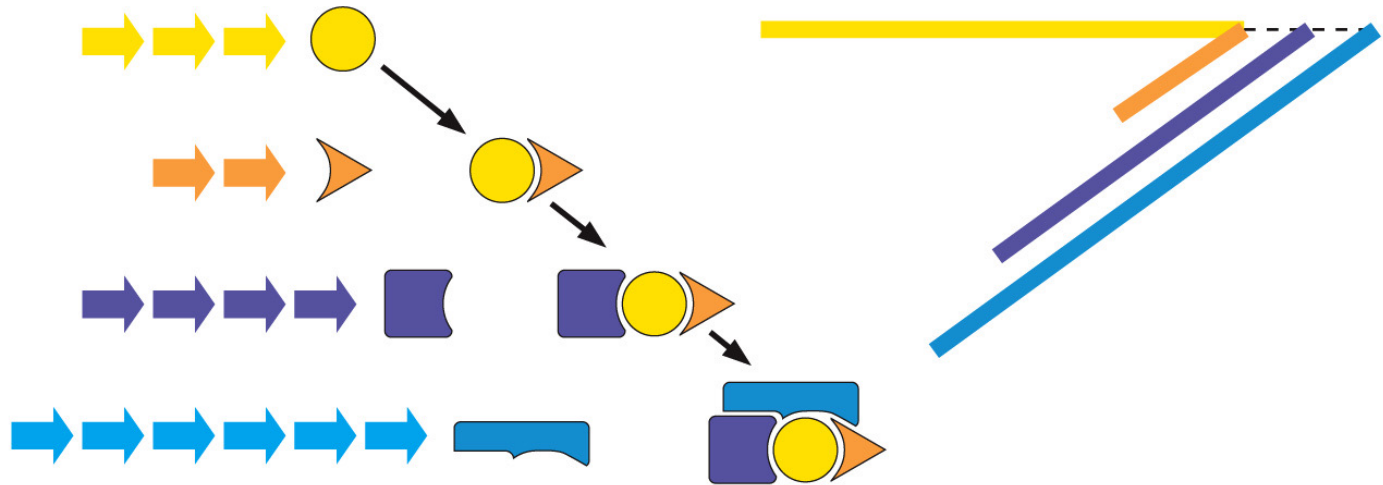


Figure 8.2a Molecular Biology of Assemblies and Machines (© Garland Science 2016)

# bacteriophage T4

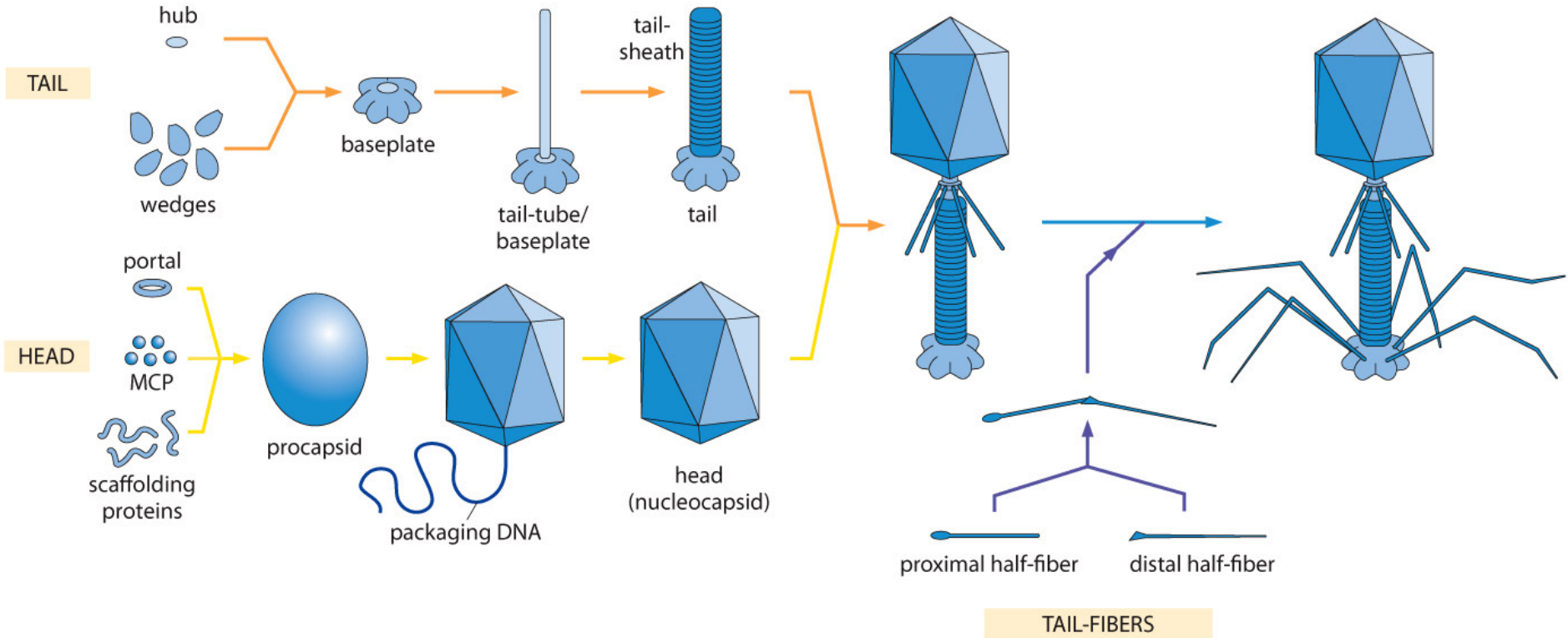


Figure 8.2b Molecular Biology of Assemblies and Machines (© Garland Science 2016)

# recognition

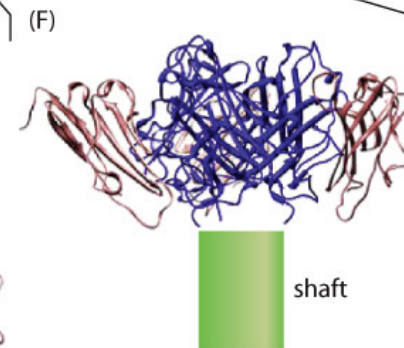
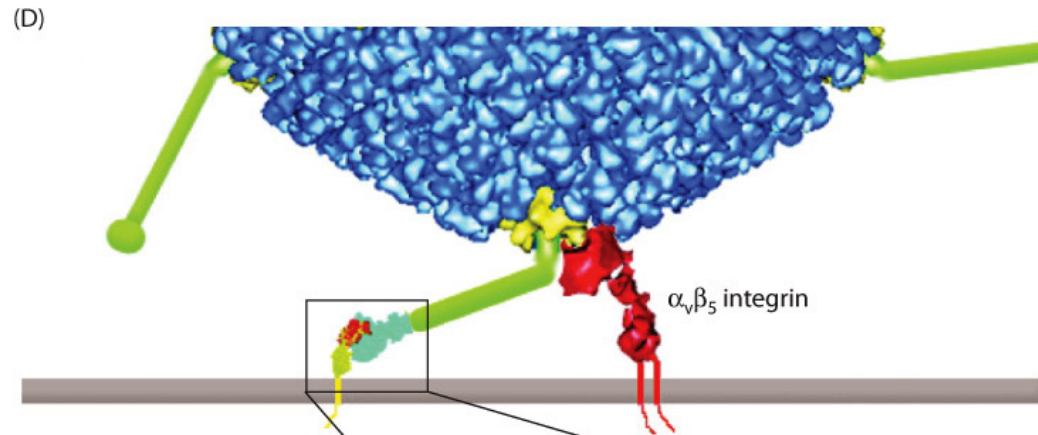
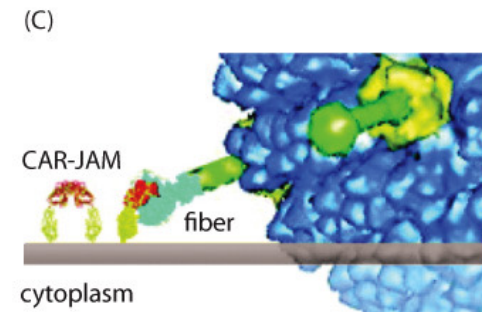
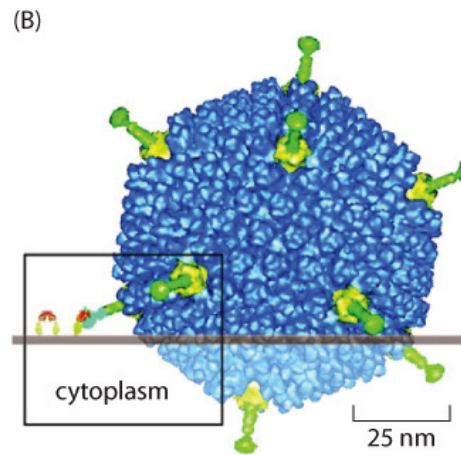
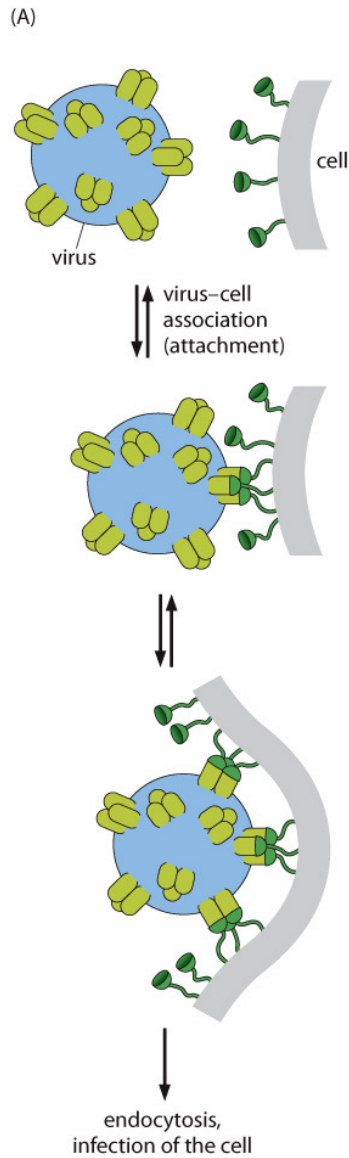
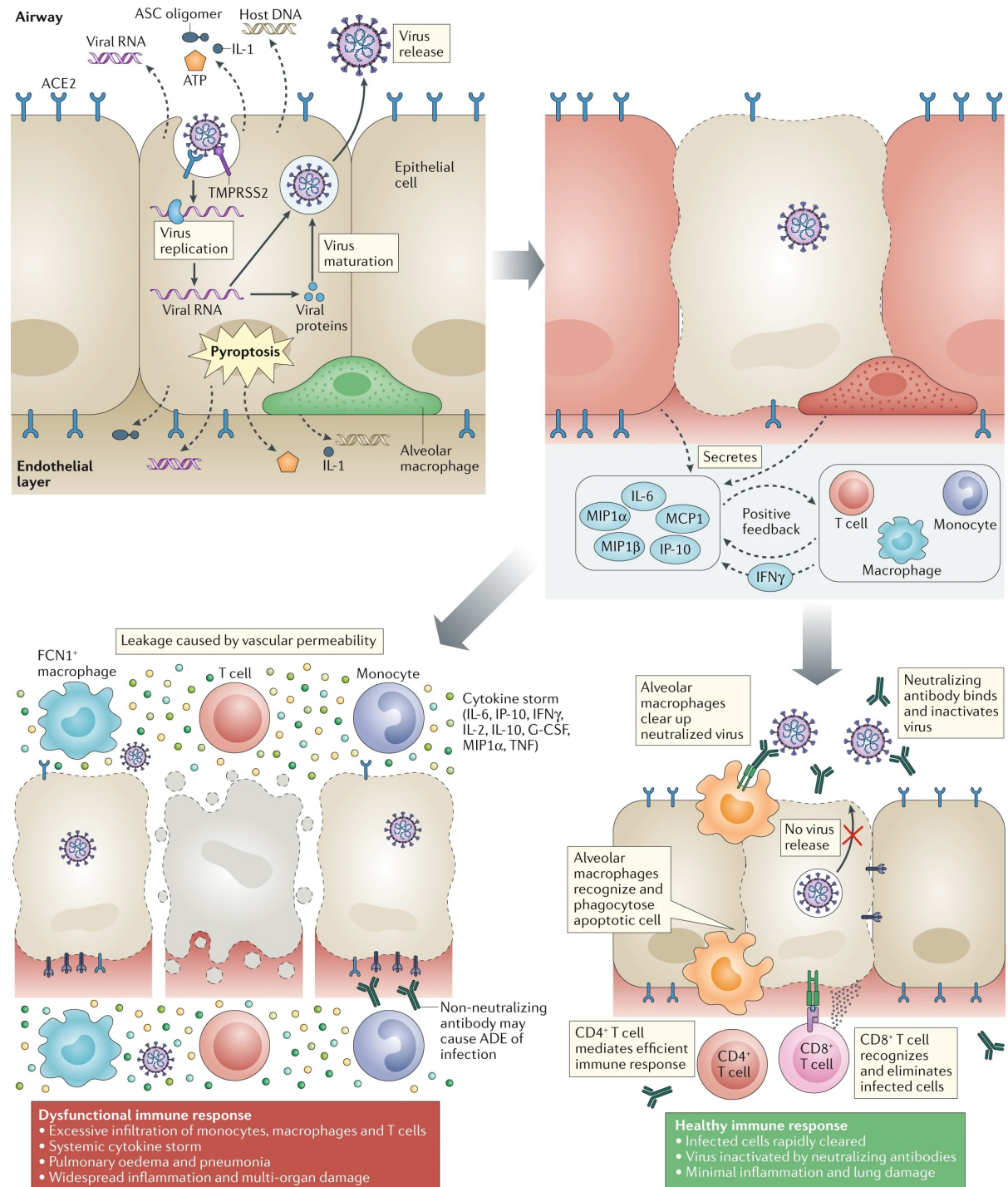


Figure 8.3 Molecular Biology of Assemblies and M:

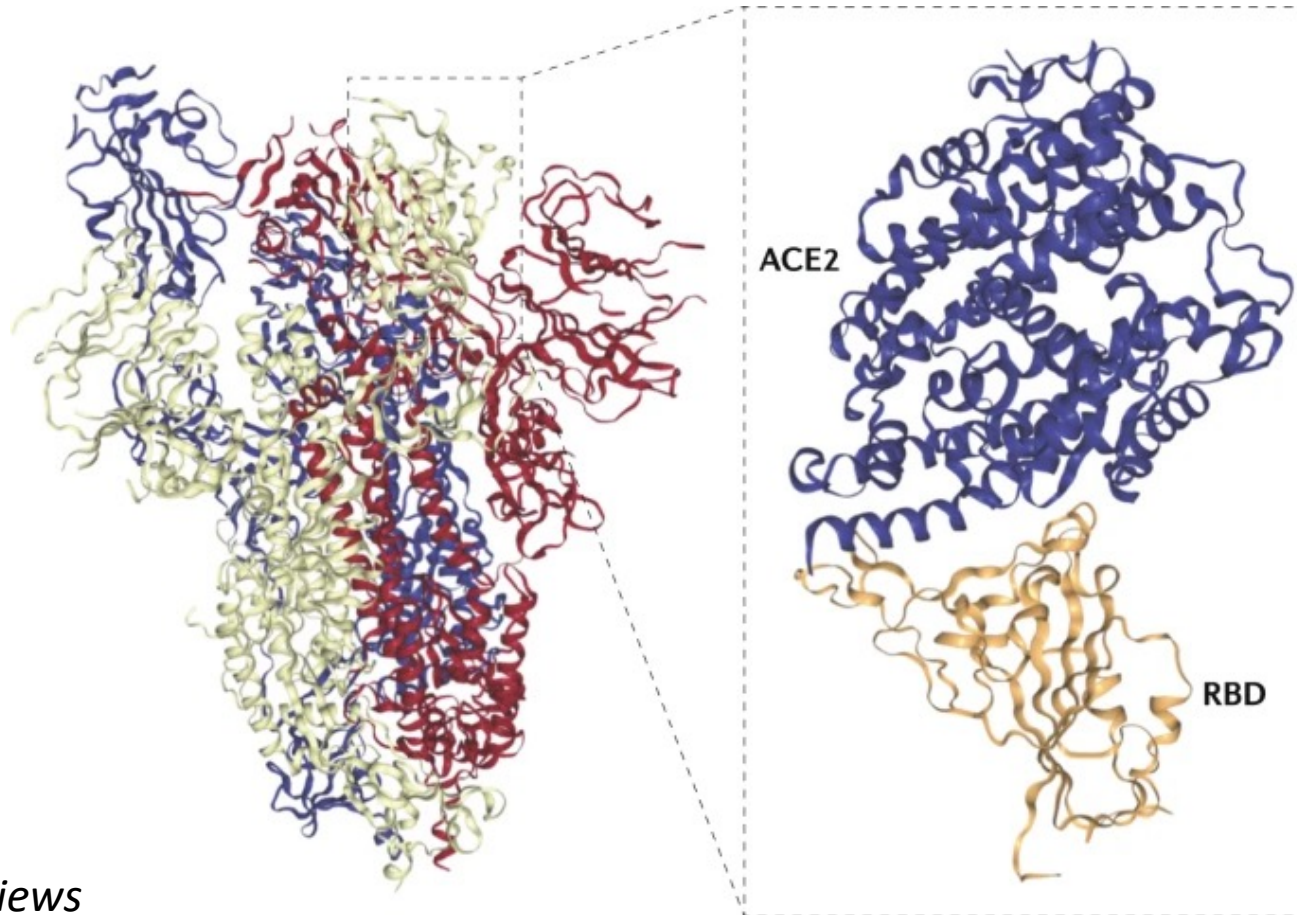


# recognition



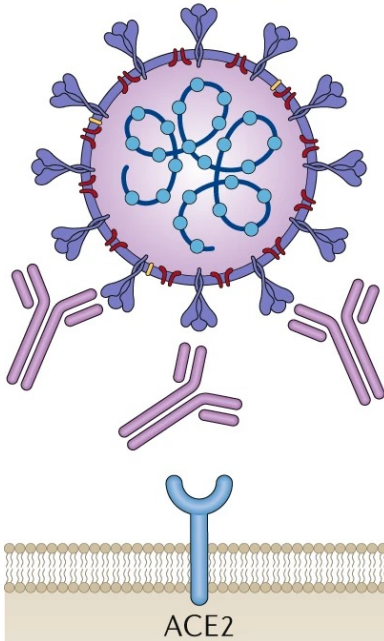
*Nature Reviews Immunology* volume 20, pages 363–374 (2020)

# The structure of the trimeric spike protein of SARS-CoV-2.

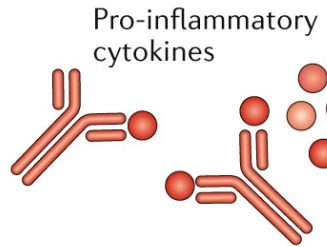


# Potential therapeutic approaches

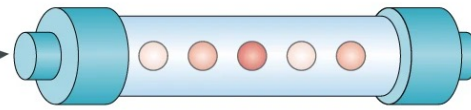
**1**  
Antibodies to spike resulting from vaccination or adoptive transfer block virus binding to ACE2



**4**



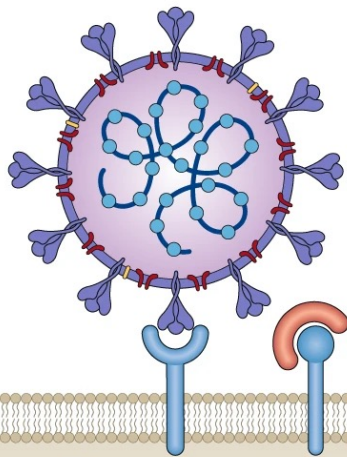
Mechanical filtration



Pro-inflammatory cytokines can be neutralized through transfer of blocking antibodies, or through mechanical removal from the blood

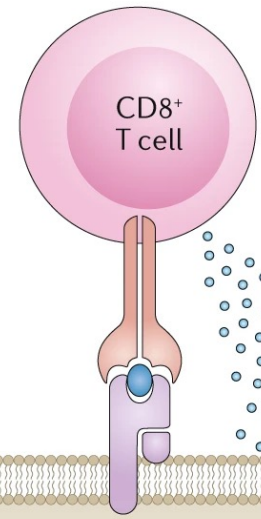
**3**

Virus-specific effector CD8<sup>+</sup> T cells arising from vaccinations recognize infected cells, secreting cytotoxic granules to kill infected cells



**2**

Protease inhibitor blocks and prevents TMRSS2 from activating the spike protein through protease cleavage

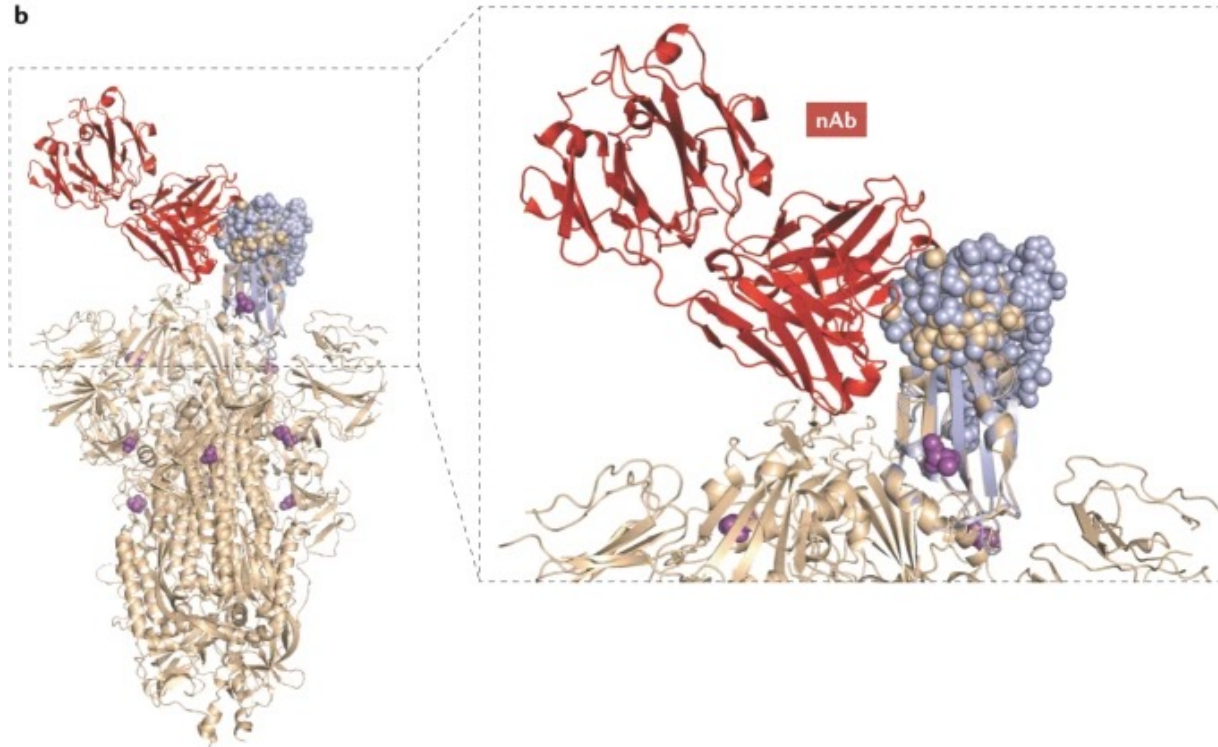


TMRSS2



**a**

SARS-CoV spike protein (316–510)	316	FPNITNLCPPFGEVFNATKFP	SVYAWERKKISNCVADYSVLYNST	TF	FFSTFK	365
SARS-CoV-2 spike protein (338–533)	338	FPNITNLCPPFGEVFNATRFAS	SVYAWNRKRISNCVADYSVLYNSA	S	FFSTFK	387
SARS-CoV spike protein (316–510)	366	CYGVSA	TKLNDLCFSNVYADSFVVKGDD	VVRQIAPGQTGVIADYNYKLPDD	415	
SARS-CoV-2 spike protein (338–533)	388	CYGVSP	TKLNDLCFTNVYADSFVIRGDE	VVRQIAPGQTGKIADYNYKLPDD	437	
SARS-CoV spike protein (316–510)	416	FMGCVLAWNTRNIDATST	GNVNYKYRYLRHGKLRPFERDIS	NV	FFSPDGK	465
SARS-CoV-2 spike protein (338–533)	438	FTGCVIAWNSNLD	SKVGGNINYLYRFRKSNLKP	PFERDISTE	YQAGST	487
SARS-CoV spike protein (316–510)	466	PCT-PPALNCYWPLNDYGFY	TTTGIGYQPYRVVLSFELLNAP	ATV	510	
SARS-CoV-2 spike protein (338–533)	488	PCNGVEGFNCYFP	LQSYGFQPTNGVGYQPYRVVLSFELL	HAP	ATV 533	

**b**

# History

- In 1953, Crick & Watson proposed ... principles of virus structure
  - Key insight:
    - Limited volume of virion capsid => nucleic acid sufficient to code for only a few sorts of proteins of limited size
  - Conclusion:
    - Identical subunits in identical environments
    - Icosahedral, dodecahedral symmetry
- In 50's & 60's Klug and others confirmed that several (unrelated) “spherical” viruses had icosahedral symmetry
  - (Used negative staining & electron microscopy)
- Conclusion:
  - Icosahedral symmetry is preferred in virus structure



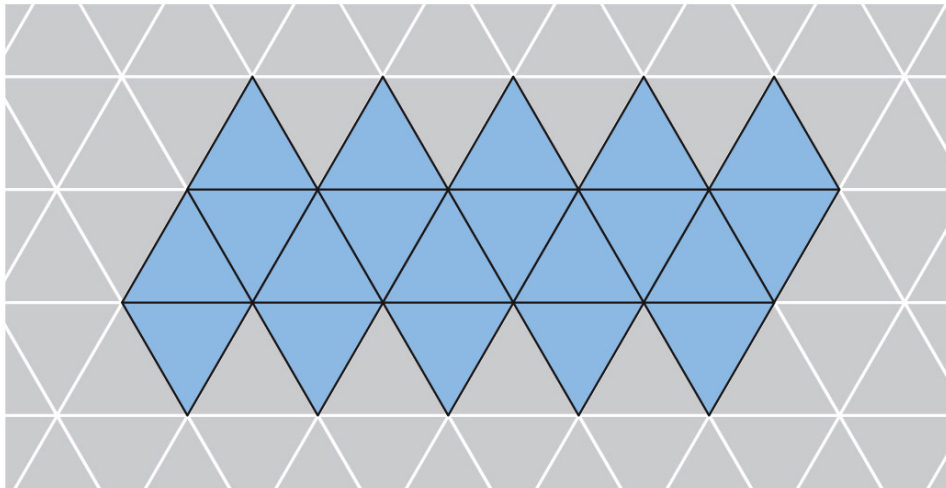
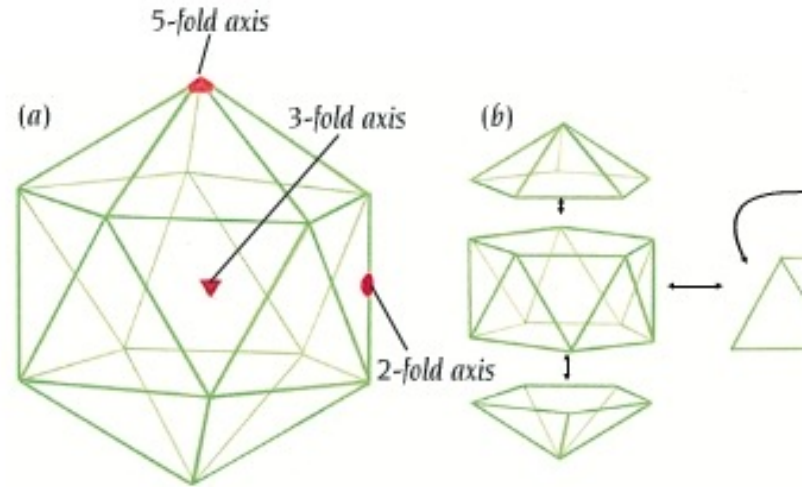
# Icosahedral Symmetry

12 vertices

20 faces (equilateral triangles)

5-3-2 symmetry axes

60 identical\* subunits  
in identical environments  
can form icosahedral shell  
\* asymmetric



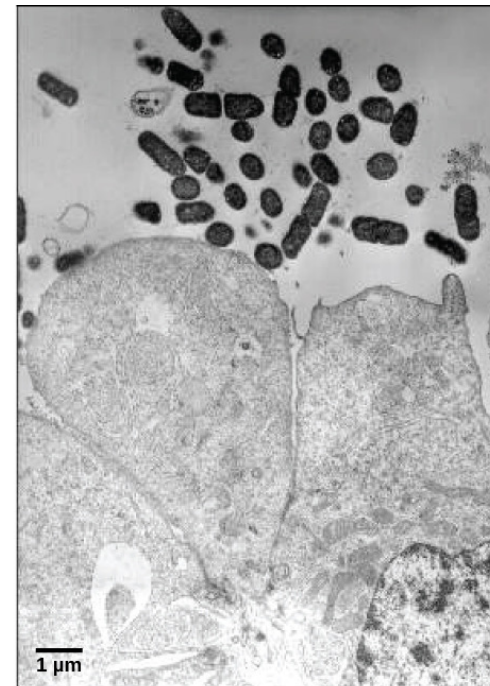
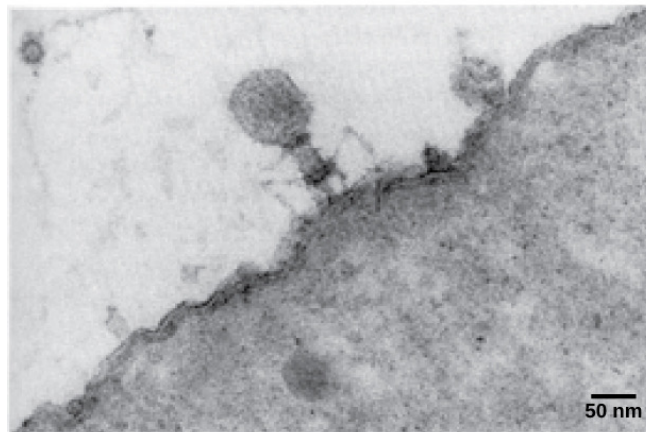
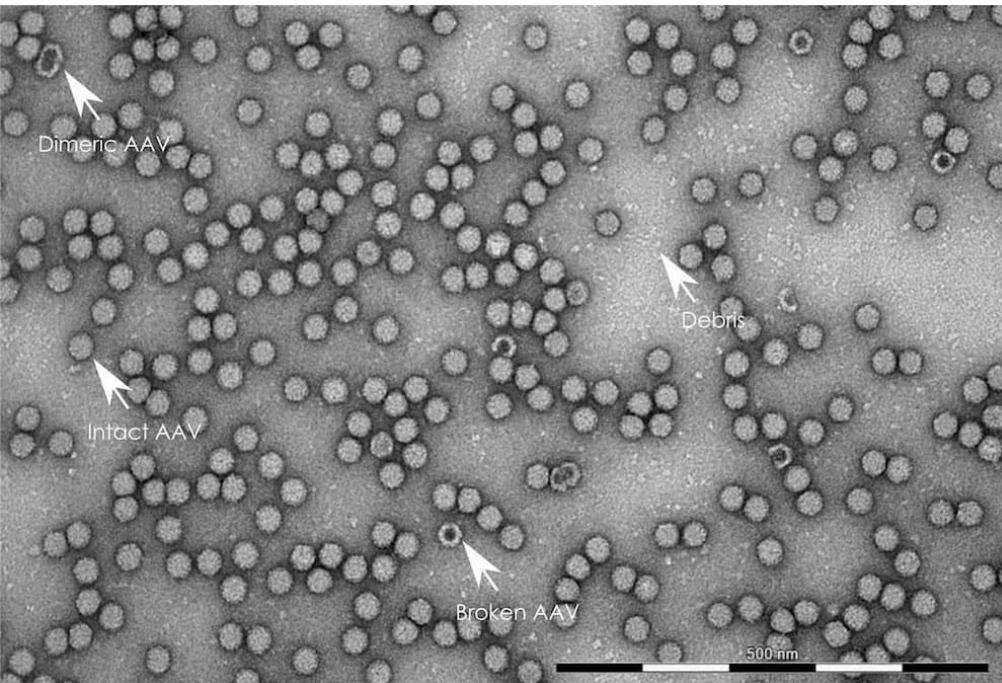
# But ...

- Clear evolutionary pressure to make larger capsid
  - Using larger subunits helps very little
  - Using more subunits helps a lot
- Not possible to form icosahedral shell (of identical units in identical environments) with more than 60 subunits
- Viruses with more than 60 subunits were observed
- In 1962, Caspar & Klug proposed the theory of “quasi-equivalence”
  - Not all protein subunits are equivalent
    - “Identical” subunits in slightly different environments
  - Only certain numbers of subunits will can be packed into closed regular lattice.

# X-ray Crystallography of Viruses

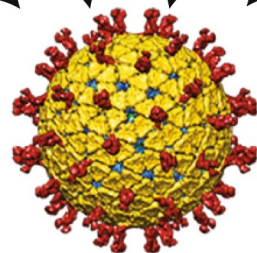
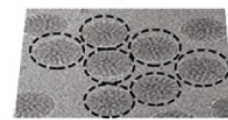
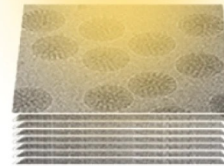
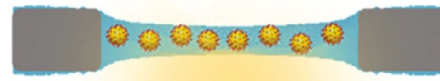
- Symmetry of protein shells makes them uniquely well-suited to crystallographic methods
- Viruses are the largest assemblies of biological macromolecules whose structures have been determined at high resolution

# Electron Microscopy

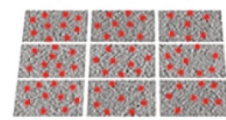
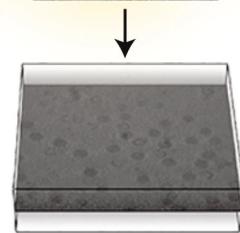
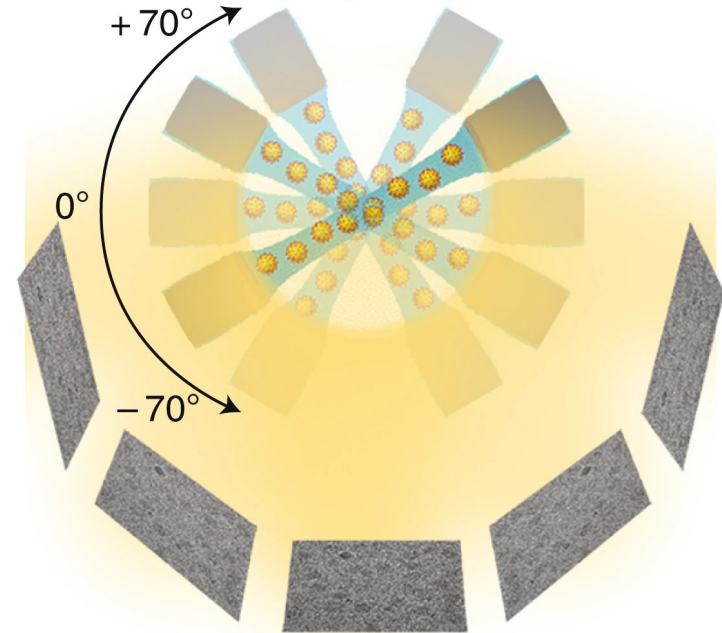


# Electron Microscopy

**a** Cryo-EM

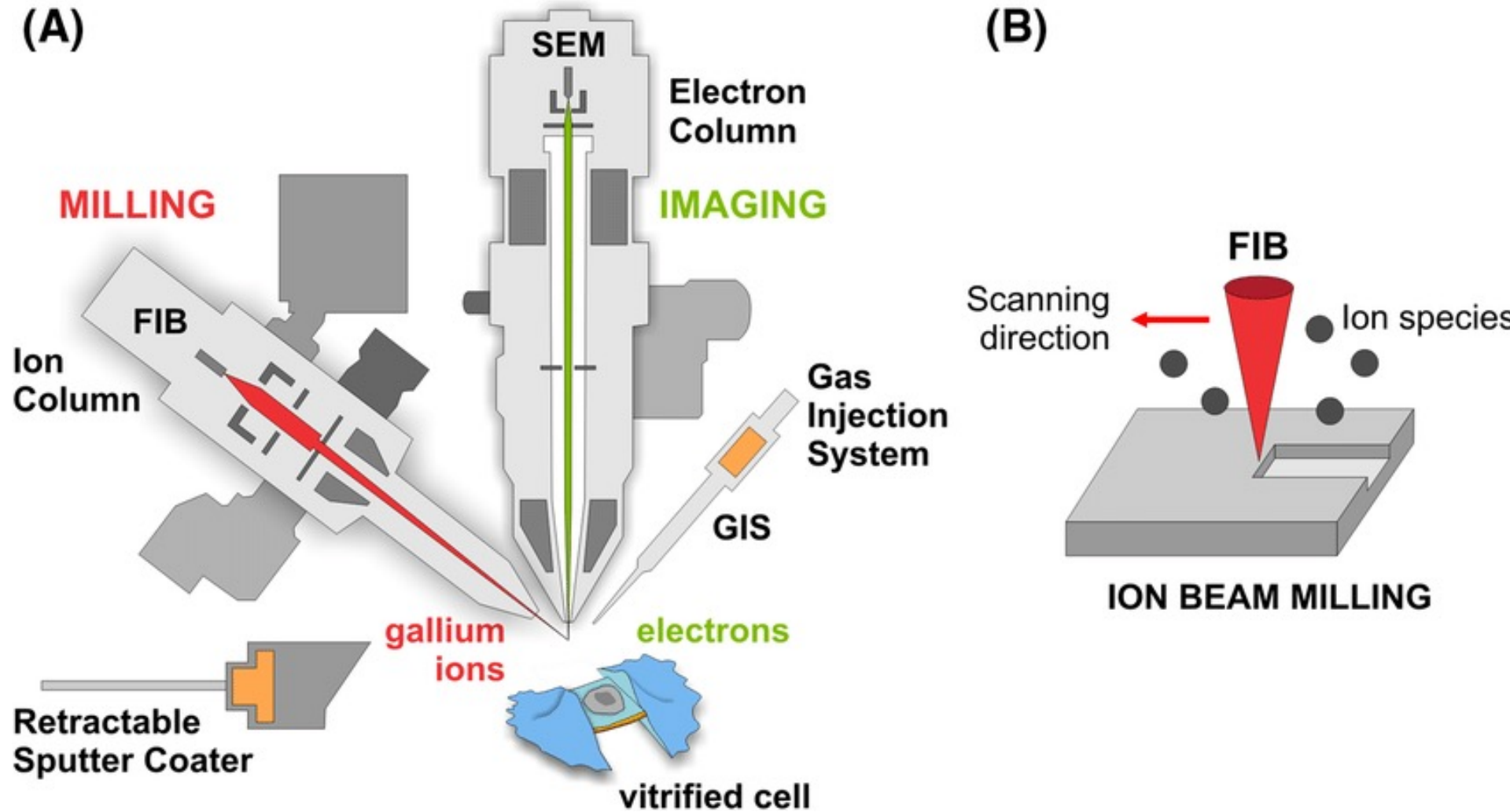


**b** Cryo-ET

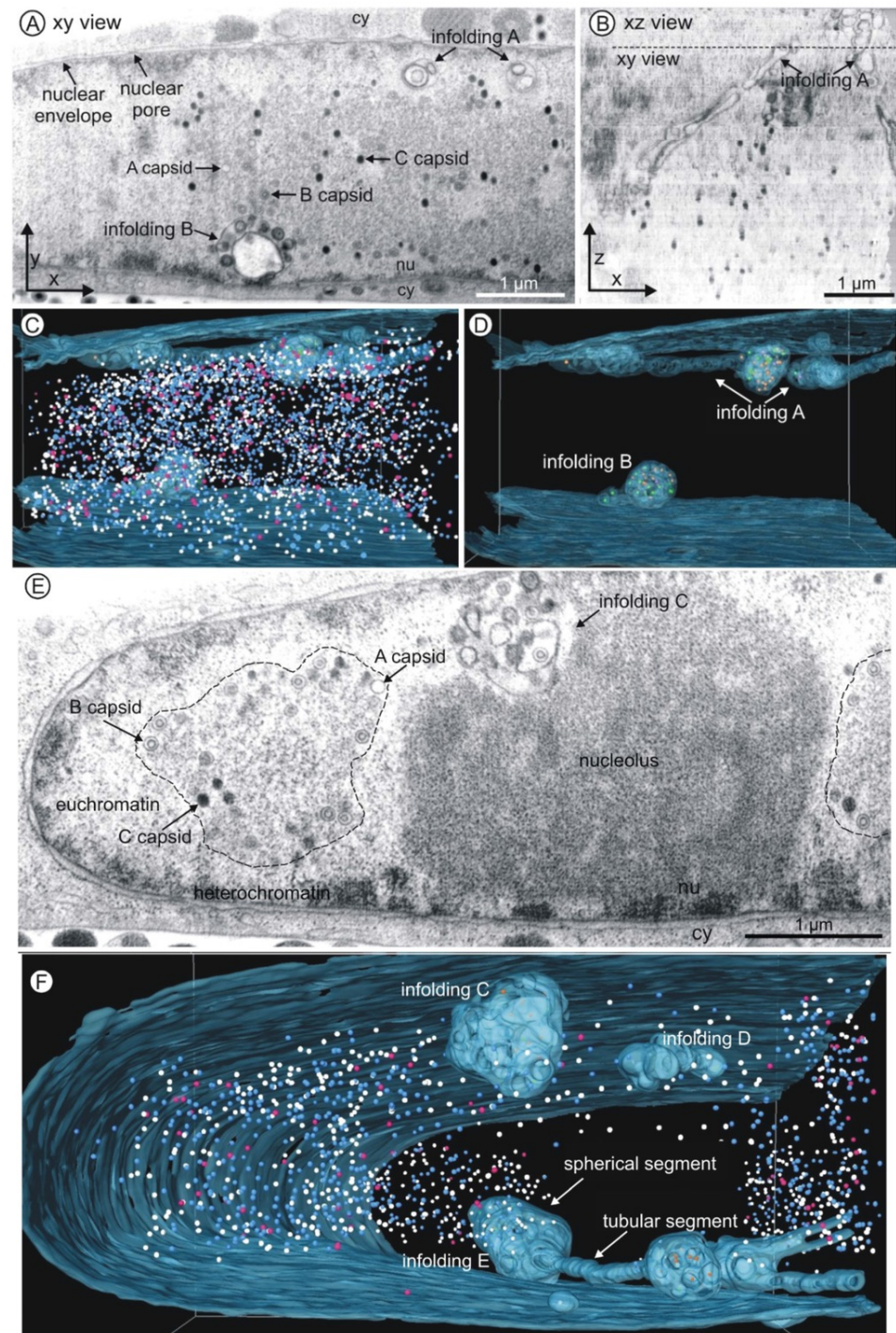




# focused ion beam

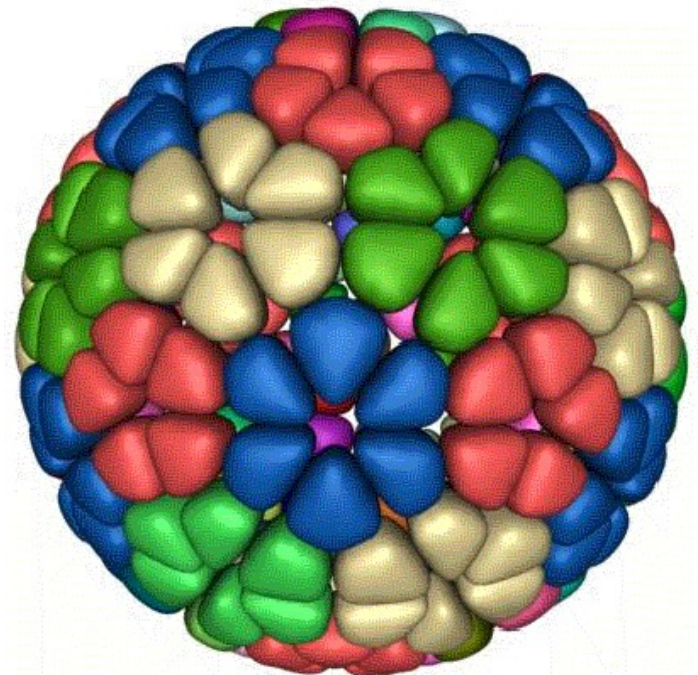
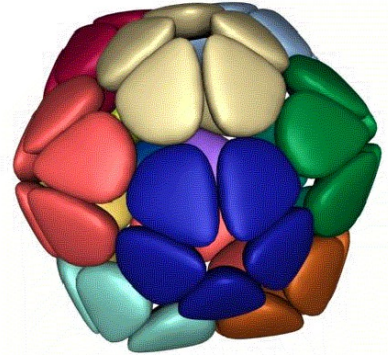


# FIB/SEM tomography of an HCMV infected nucleus



# Quasi-equivalence

- Subunits are in “minimally” different environments
  - Pentamers at vertices
  - Hexamers elsewhere
- Predicts packing arrangements of larger capsids
  - Shift from T1 to T4 packing
  - => 8-fold increase in volume





# Goldberg diagram

$$T = h^2 + hk + k^2,$$

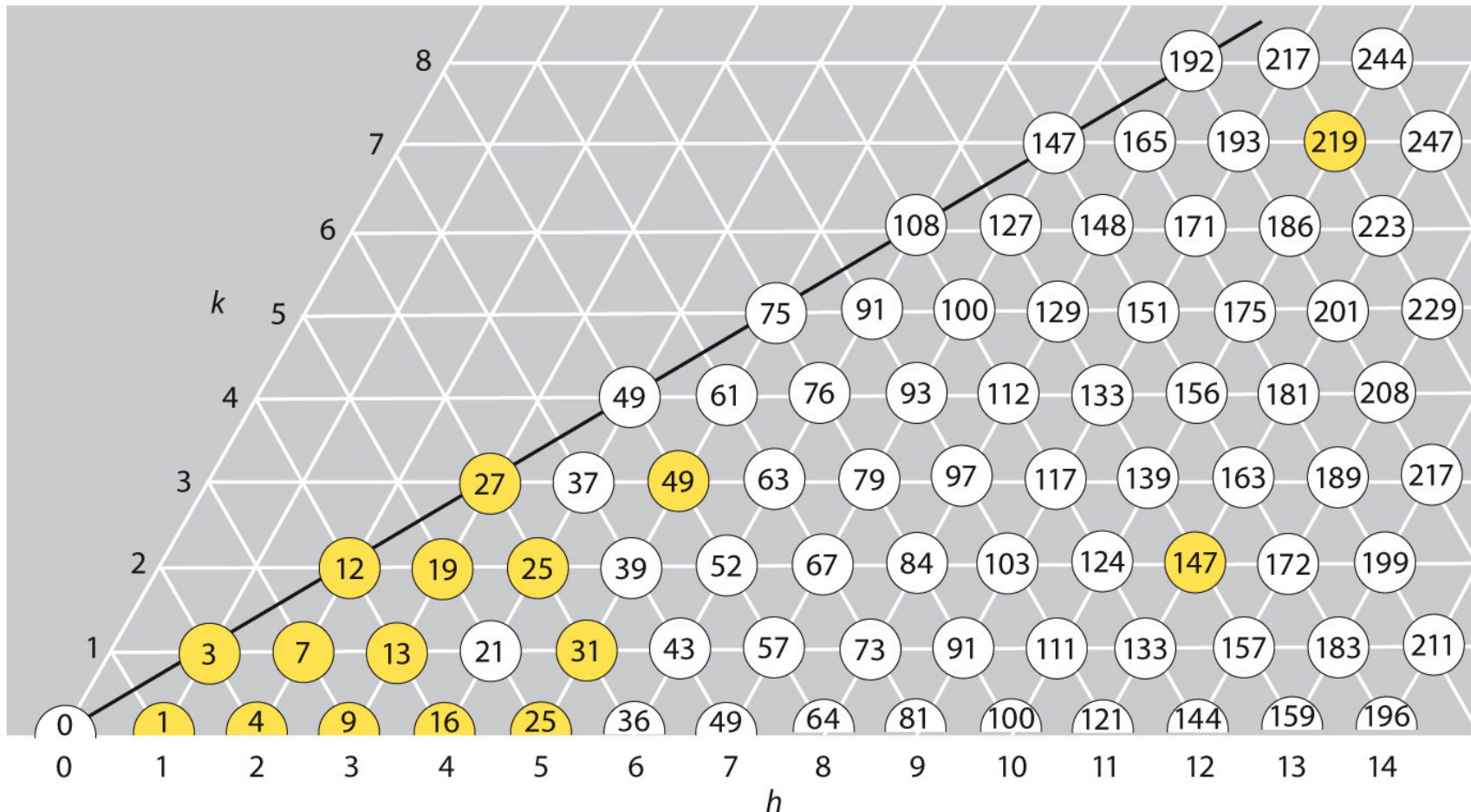
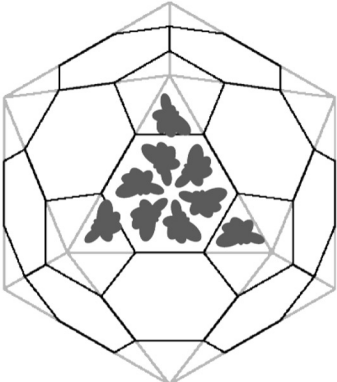


Figure 8.10 Molecular Biology of Assemblies and Machines (© Garland Science 2016)

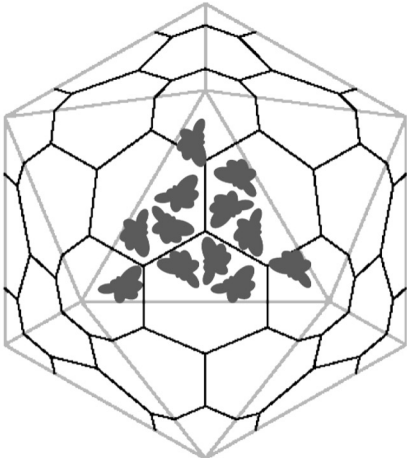
# Spherical viruses have icosahedral symmetry



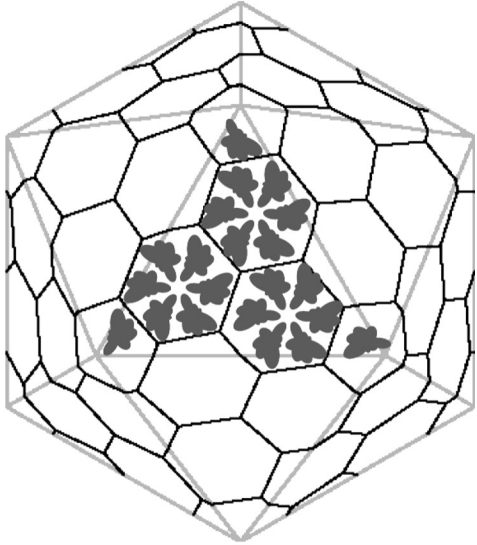
T=1



T=3



T=4

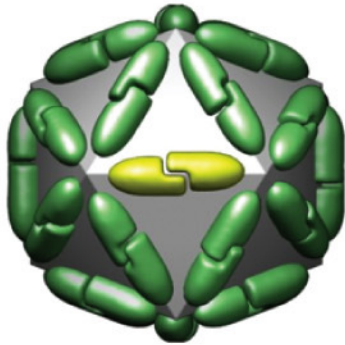


T=7

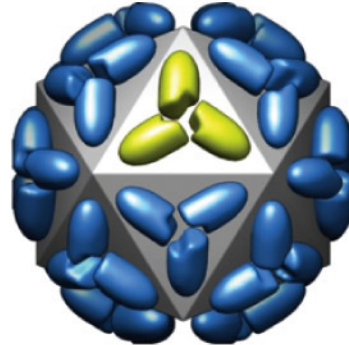


# Icosahedral capsids

(A)



(B)



(C)

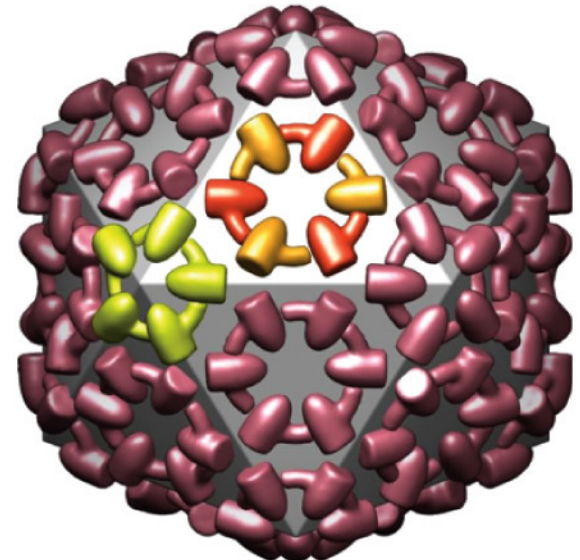
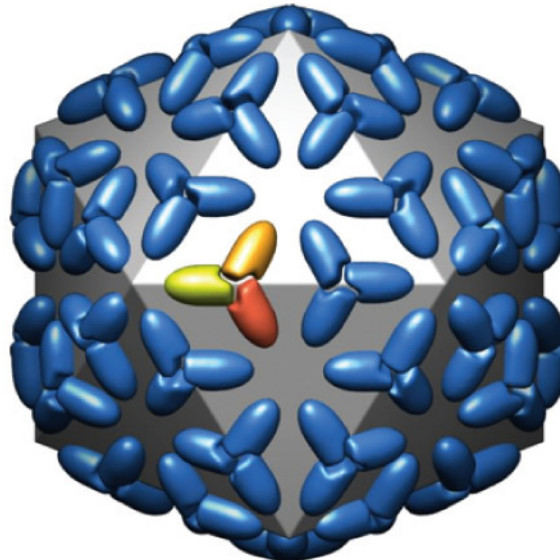
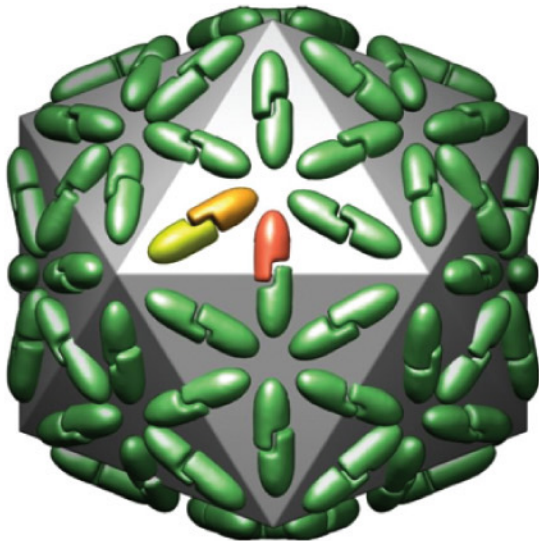
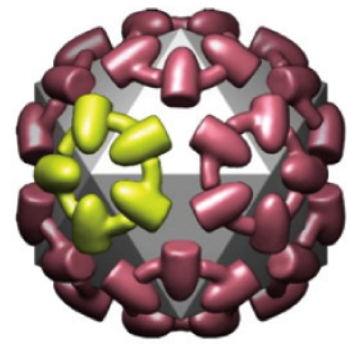


Figure 8.8 Molecular Biology of Assemblies and Machines (© Garland Science 2016)

# interactions between complementary surface patches

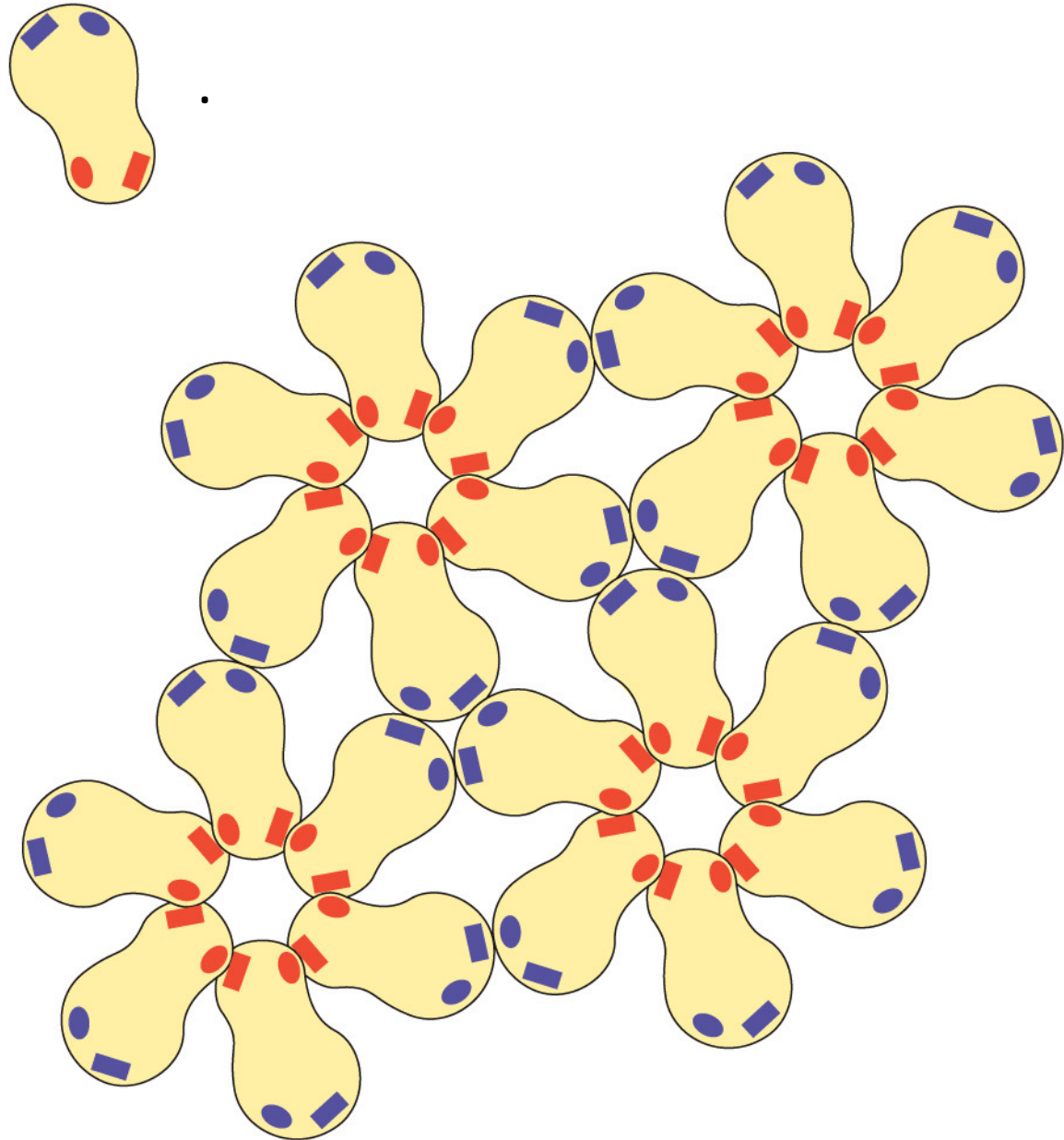


Figure 8.13 Molecular Biology of Assemblies and Machines (© Garland Science 2016)

# Helical viruses

## tobacco mosaic virus (TMV)

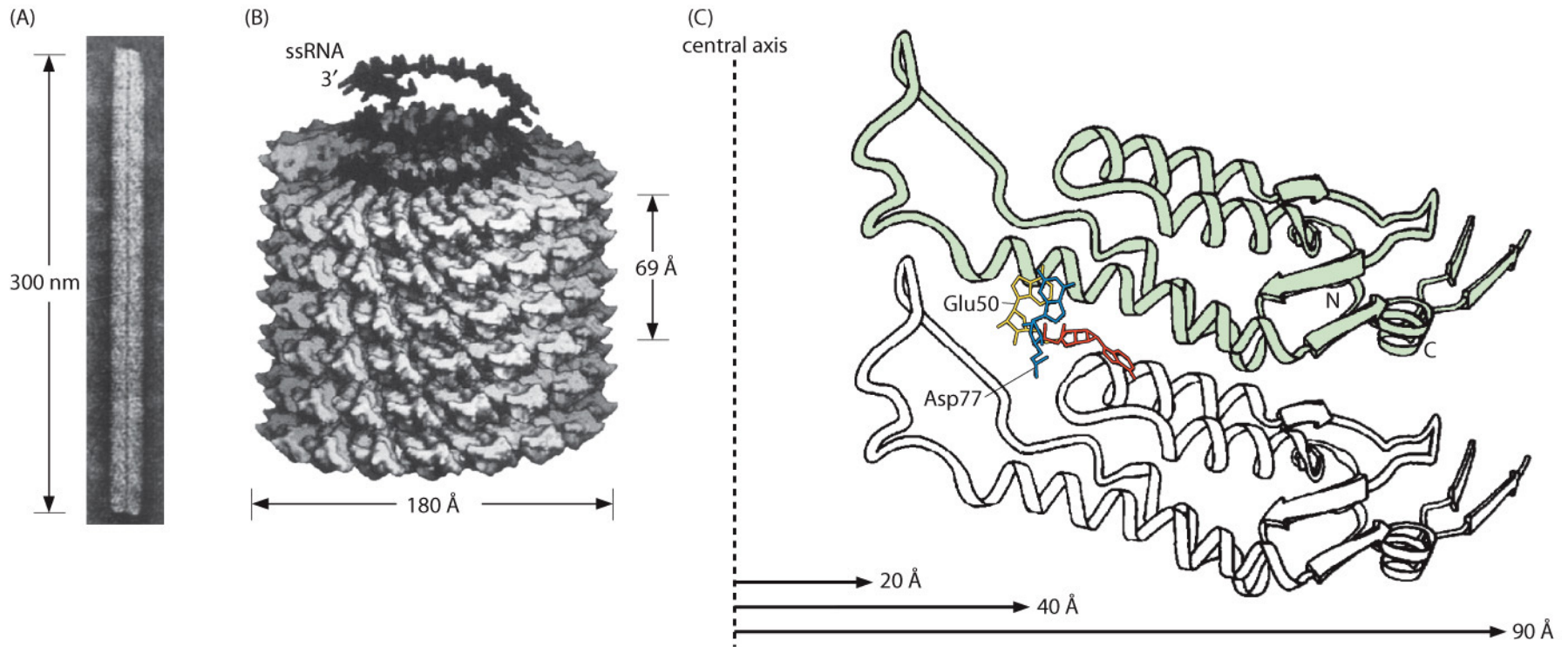
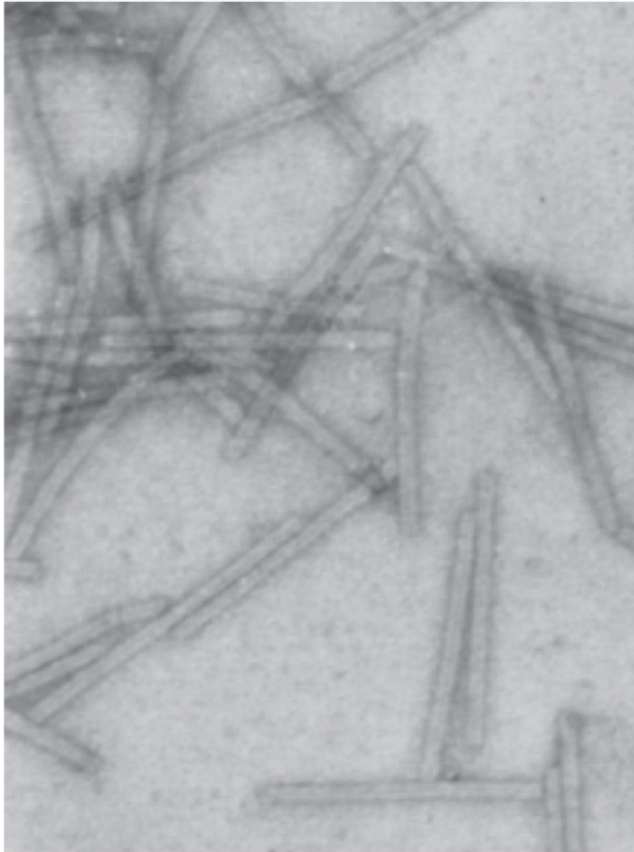


Figure 8.15 Molecular Biology of Assemblies and Machines (© Garland Science 2016)

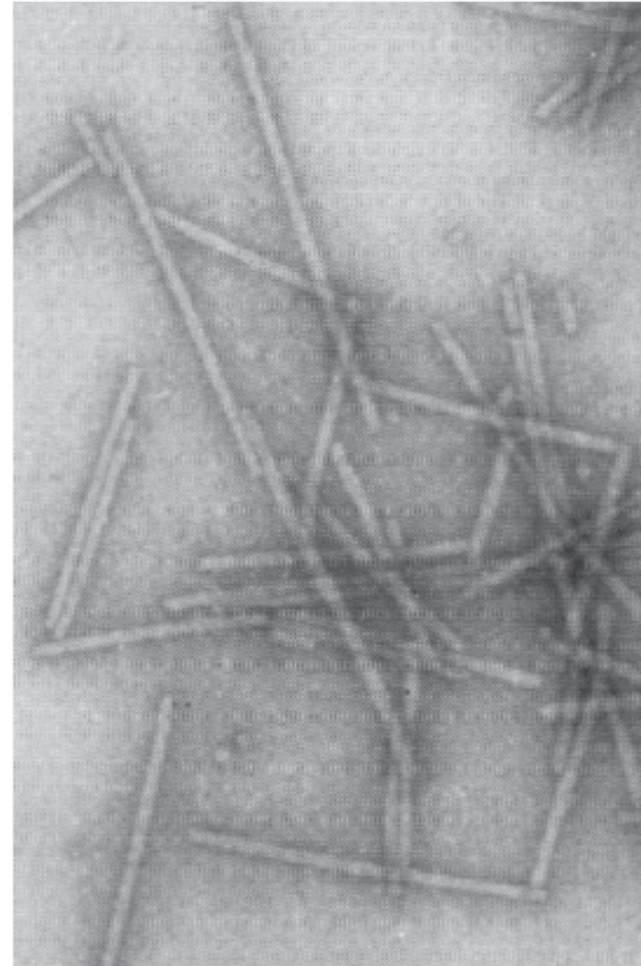


# Helical viruses

with RNA



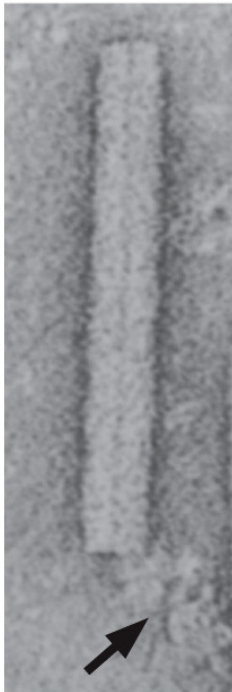
without RNA



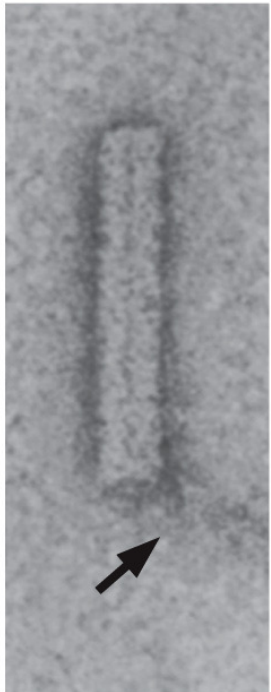
300 nm

# Disassembly of TMV

(A)



high pH



detergent

┌───┐  
25 nm

(B)



ribosome-mediated  
disassembly

Figure 8.17 Molecular Biology of Assemblies and Machines (© Garland Science 2016)



# Assembly of TMV

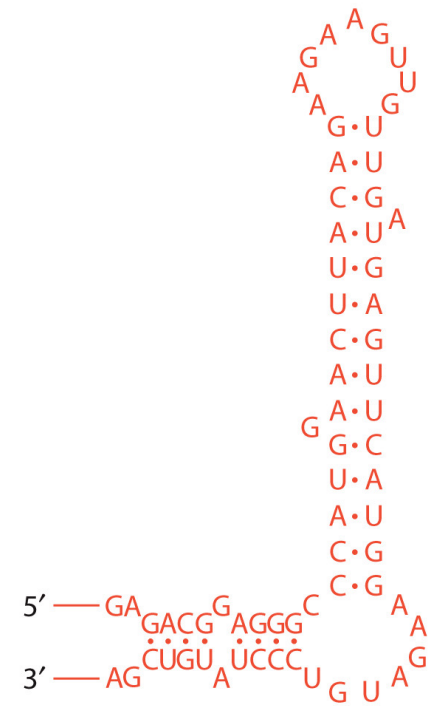
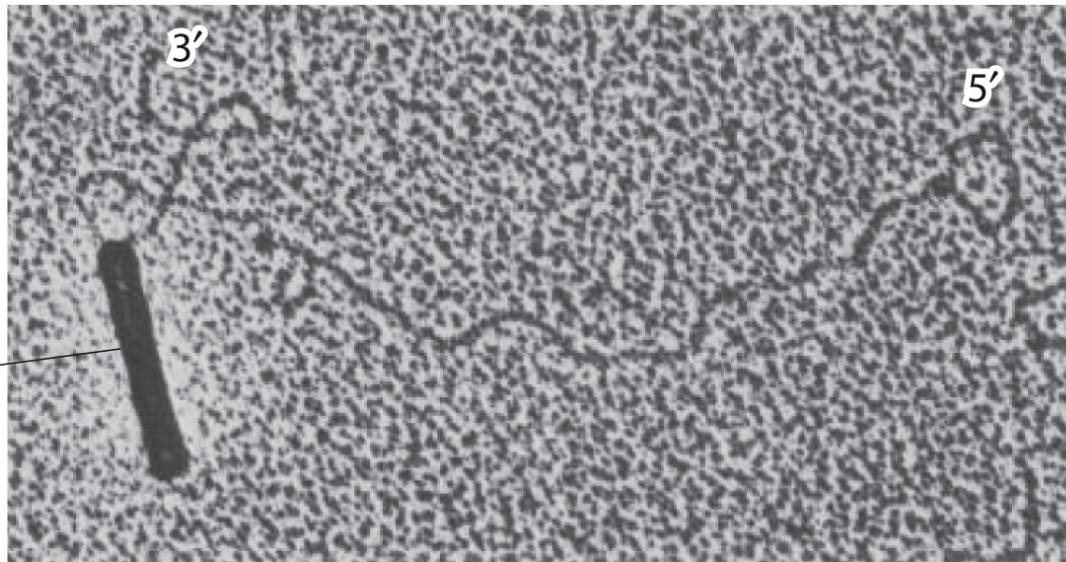


Figure 8.18a Molecular Biology of Assemblies and Machines (© Garland Science 2016)

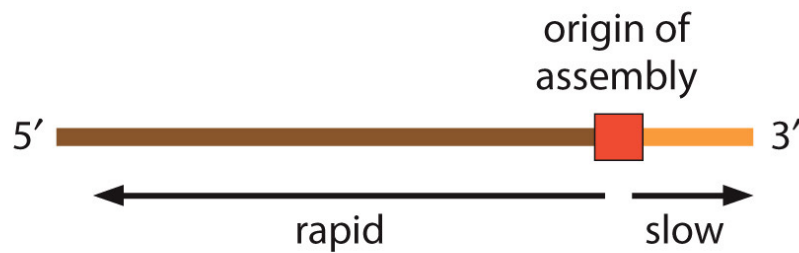


Figure 8.18b Molecular Biology of Assemblies and Machines (© Garland Science 2016)

# Assembly of TMV

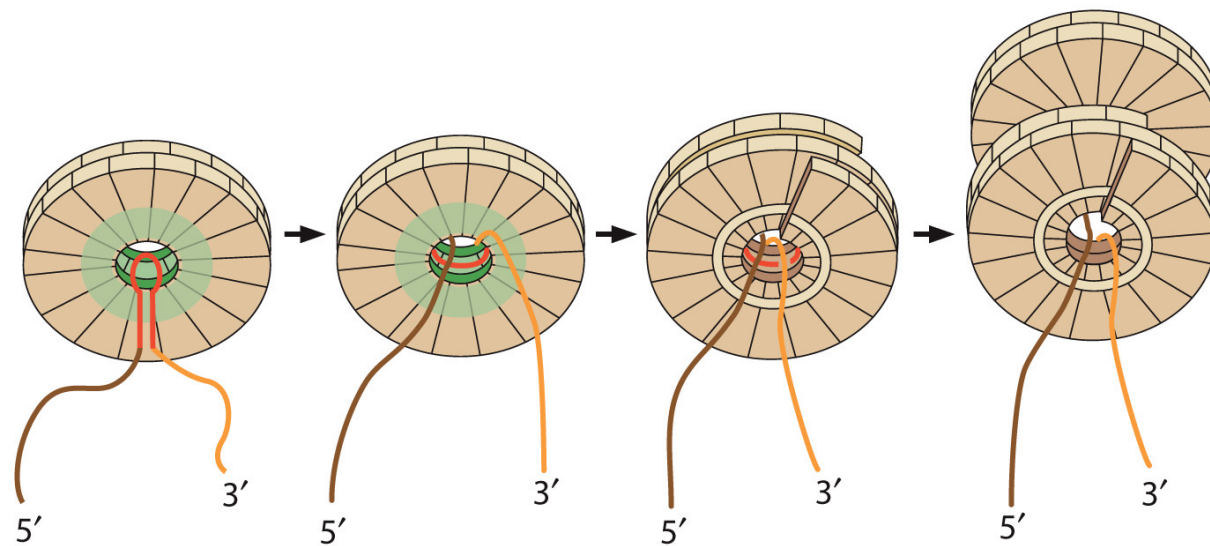


Figure 8.18c Molecular Biology of Assemblies and Machines (© Garland Science 2016)

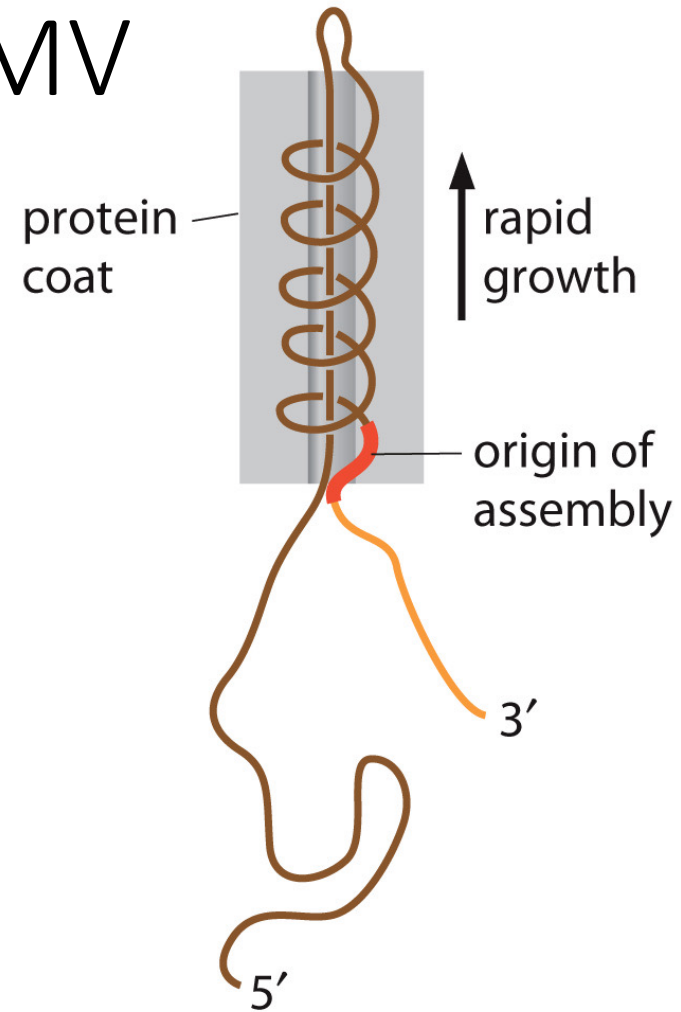


Figure 8.18d Molecular Biology of Assemblies and Machines (© Garland Science 2016)

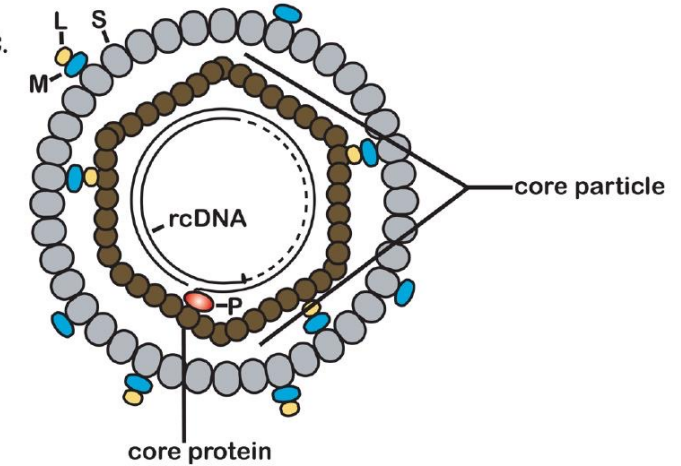
# small Icosahedral viruses

## hepatitis B virus

B.

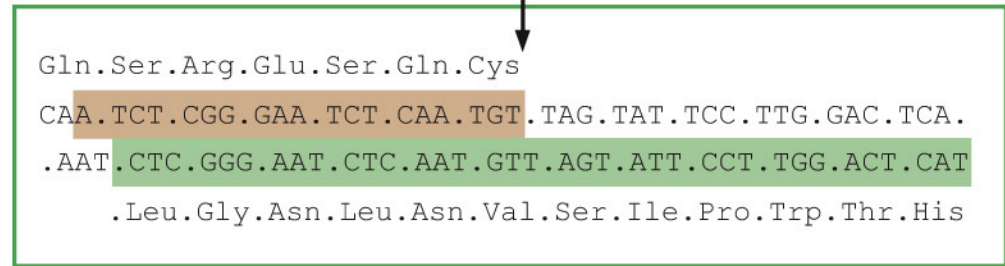


C.



(B)

end of ORF C (MCP coding sequence)



part of ORF P (polymerase coding sequence)

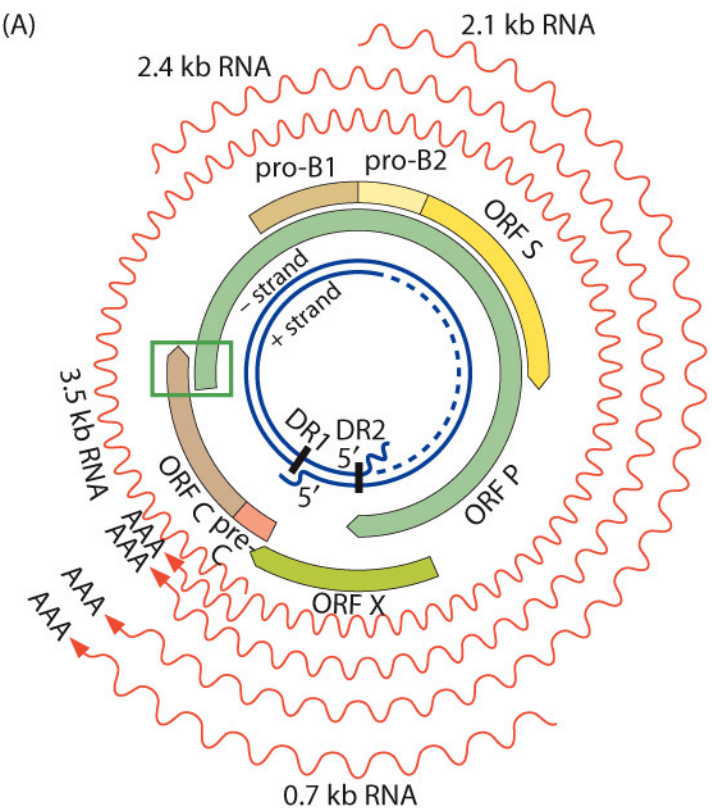
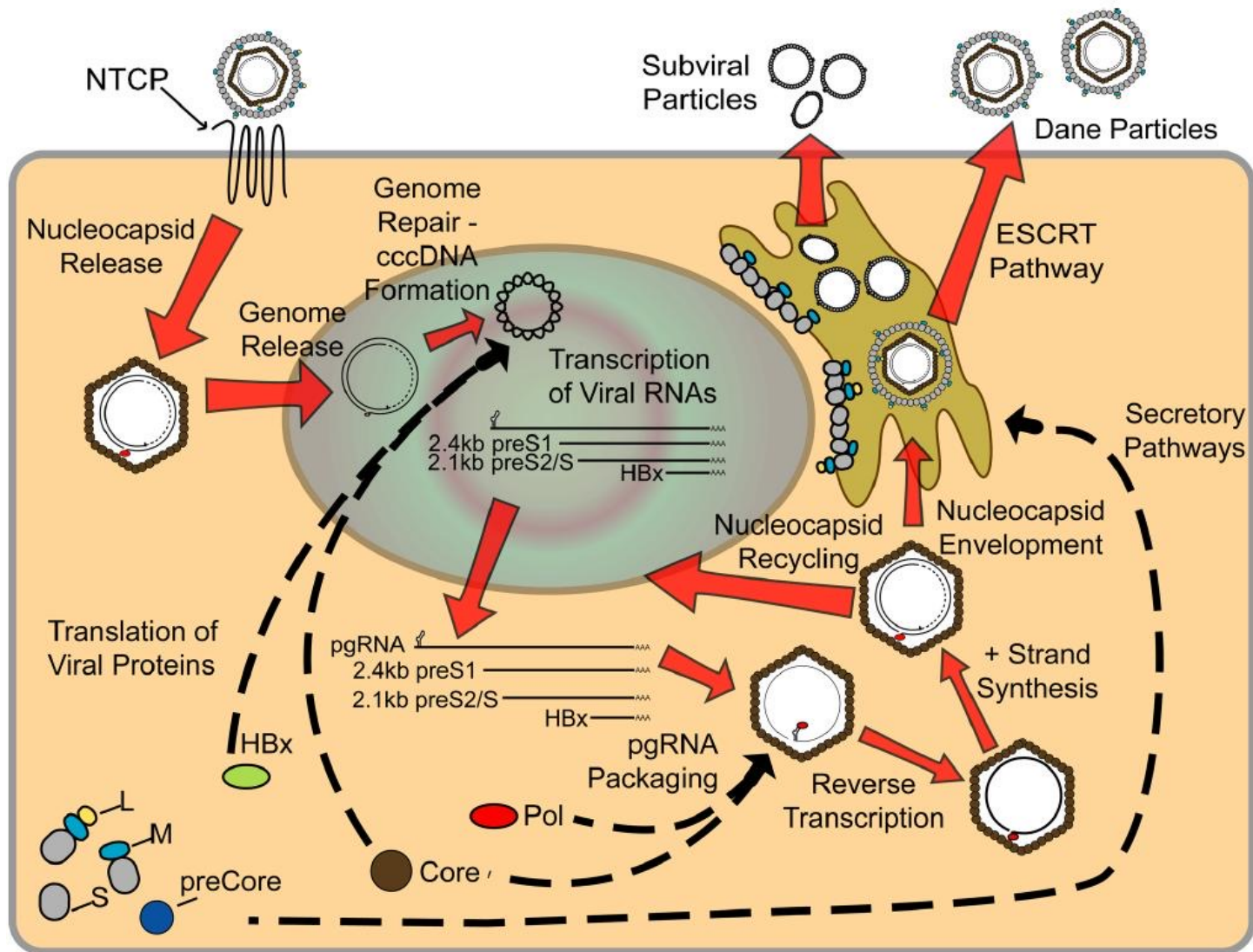


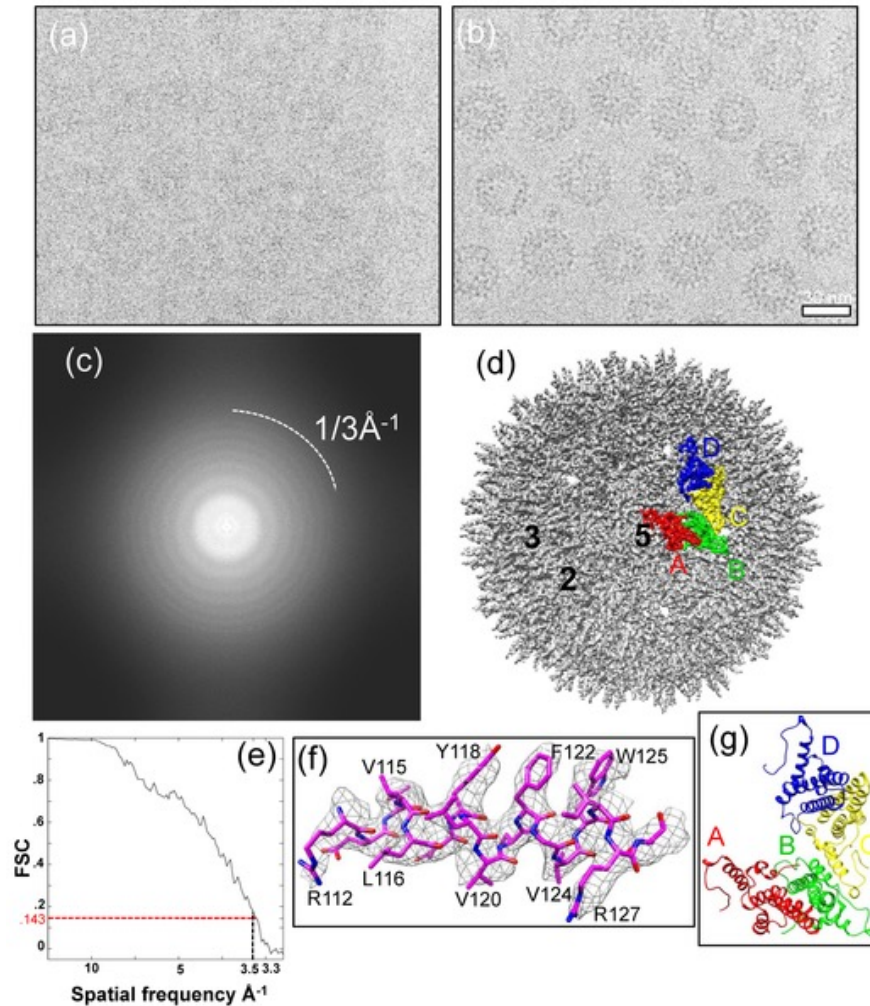
Figure 8.26 Molecular Biology of Assemblies and Machines (© Garland Science 2016)

# Life cycle





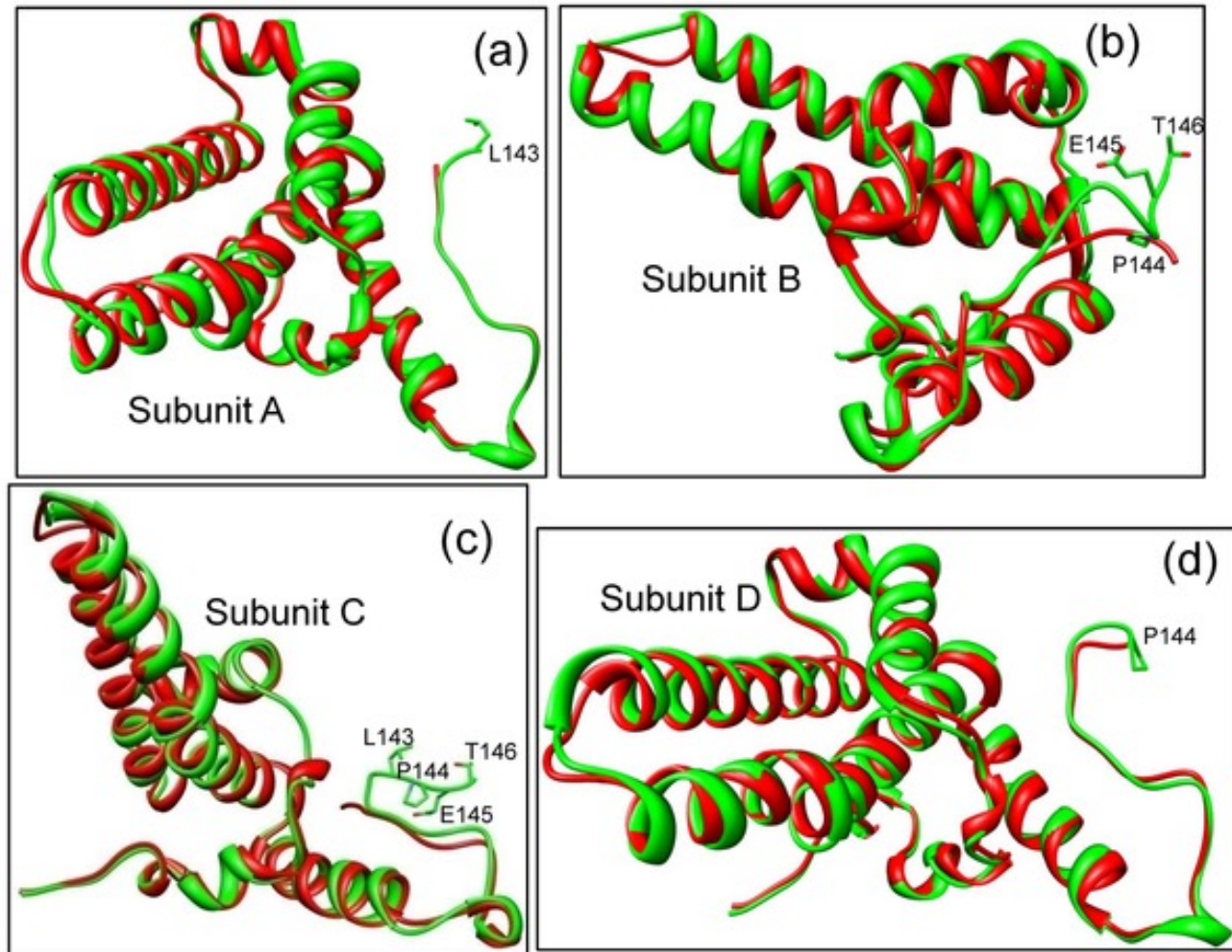
**Figure 1. CryoEM and 3D reconstruction of hepatitis B virus (HBV) core assembled from full-length HBV core proteins at 3.5Å resolution.**



Yu X, Jin L, Jih J, Shih C, Hong Zhou Z (2013) 3.5Å cryoEM Structure of Hepatitis B Virus Core Assembled from Full-Length Core Protein. PLOS ONE 8(9): e69729. <https://doi.org/10.1371/journal.pone.0069729>  
<https://journals.plos.org/plosone/article?id=10.1371/journal.pone.0069729>

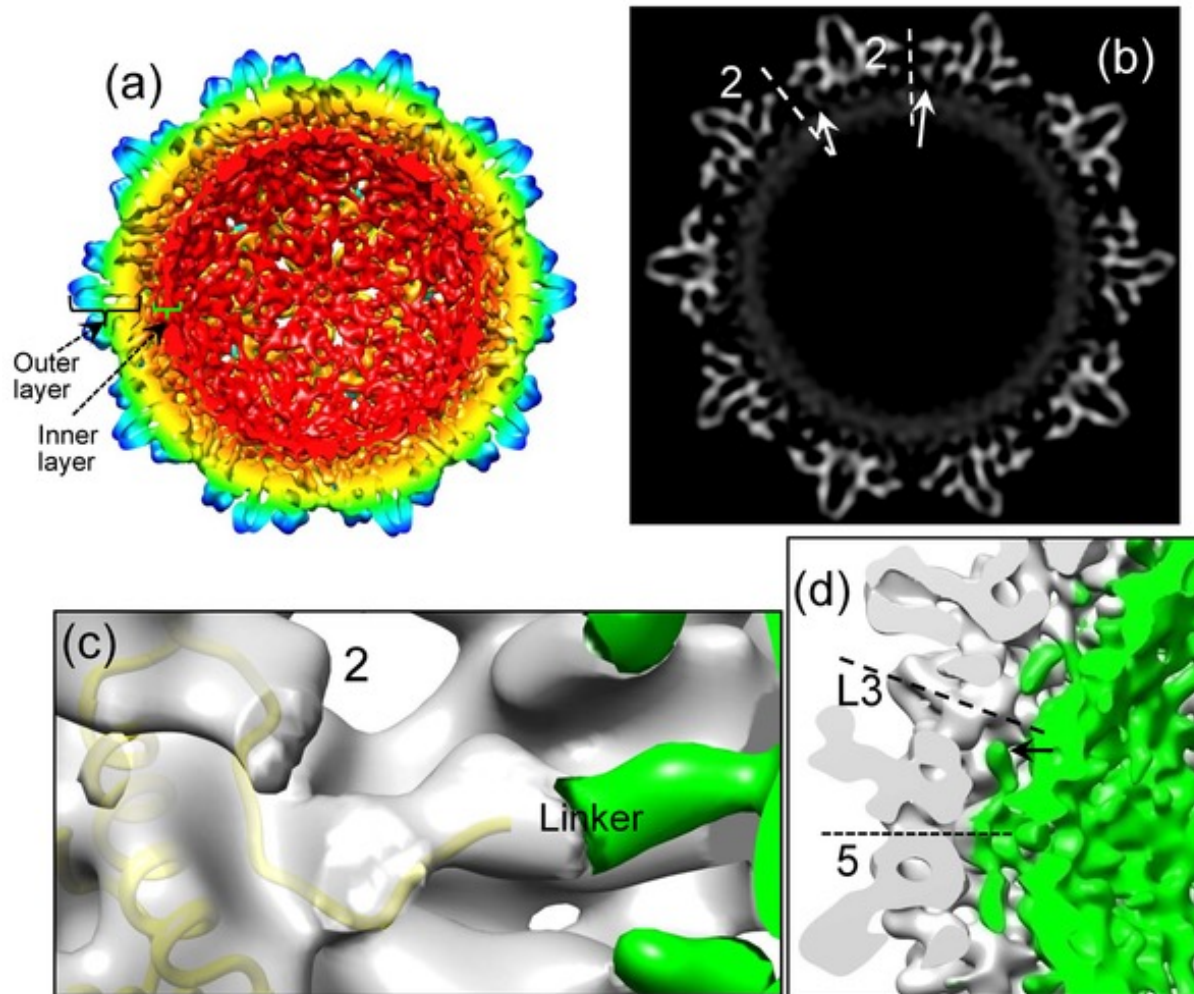


**Figure 2. Comparisons between corresponding cryoEM structures (green) and crystal structures (red) by superimposition.**



Yu X, Jin L, Jih J, Shih C, Hong Zhou Z (2013) 3.5Å cryoEM Structure of Hepatitis B Virus Core Assembled from Full-Length Core Protein. PLOS ONE 8(9): e69729. <https://doi.org/10.1371/journal.pone.0069729>  
<https://journals.plos.org/plosone/article?id=10.1371/journal.pone.0069729>

**Figure 4. Maps of HBV core reconstruction filtered to 10Å resolution.**



Yu X, Jin L, Jih J, Shih C, Hong Zhou Z (2013) 3.5Å cryoEM Structure of Hepatitis B Virus Core Assembled from Full-Length Core Protein. PLOS ONE 8(9): e69729. <https://doi.org/10.1371/journal.pone.0069729>  
<https://journals.plos.org/plosone/article?id=10.1371/journal.pone.0069729>

# neutron diffraction

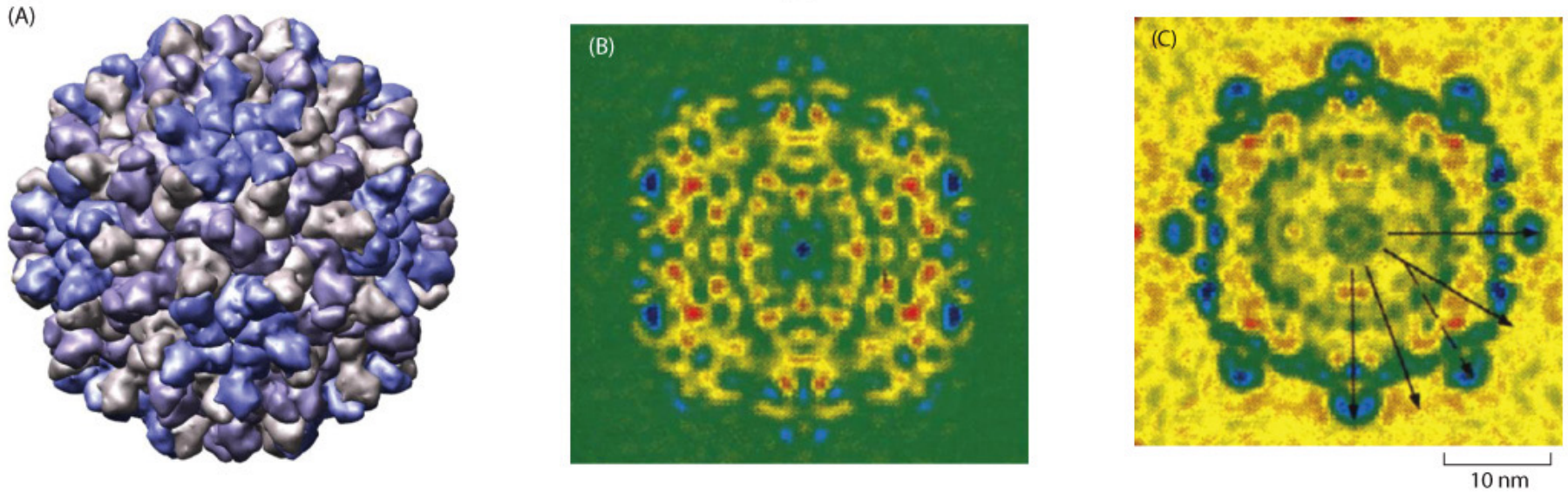
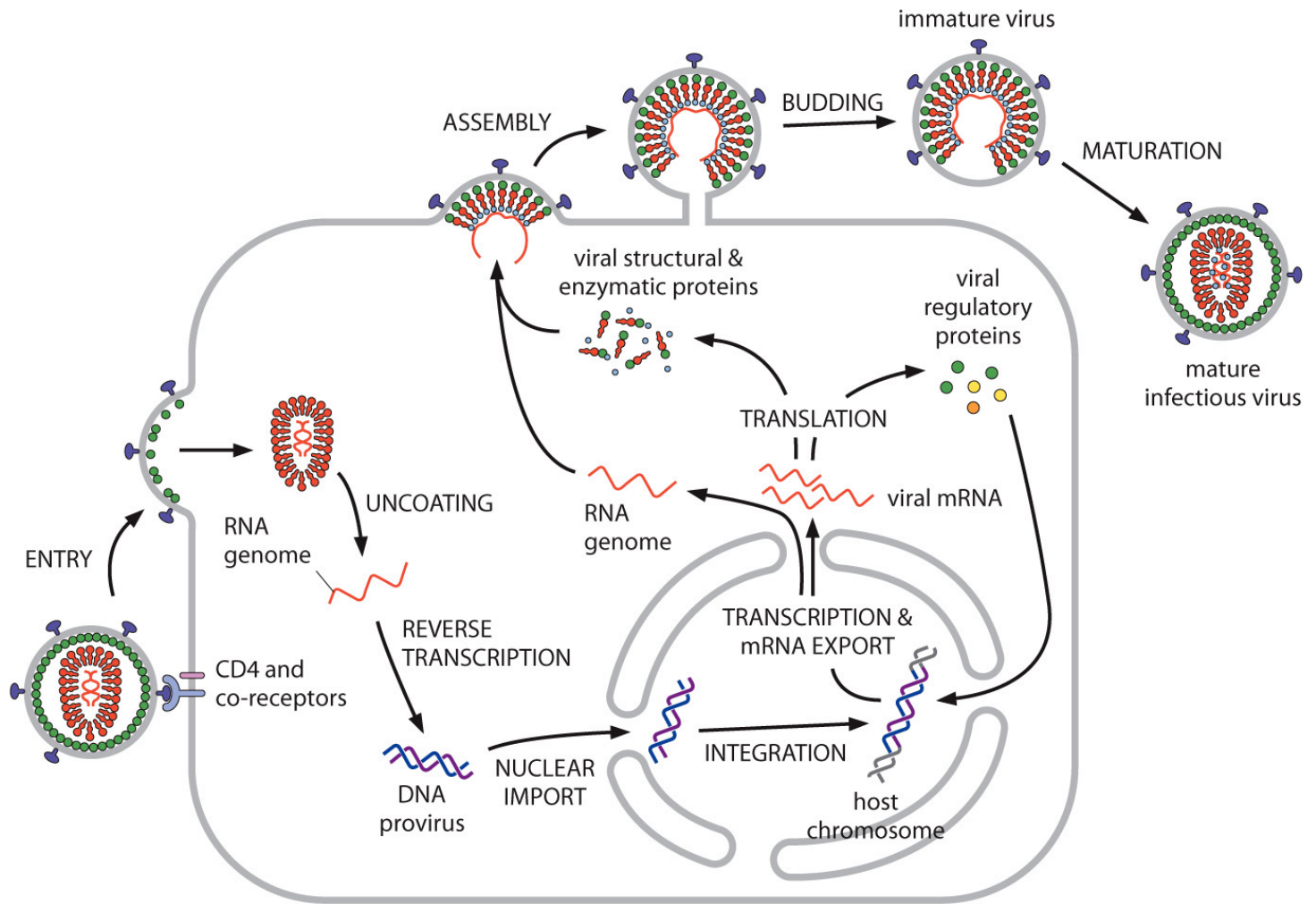


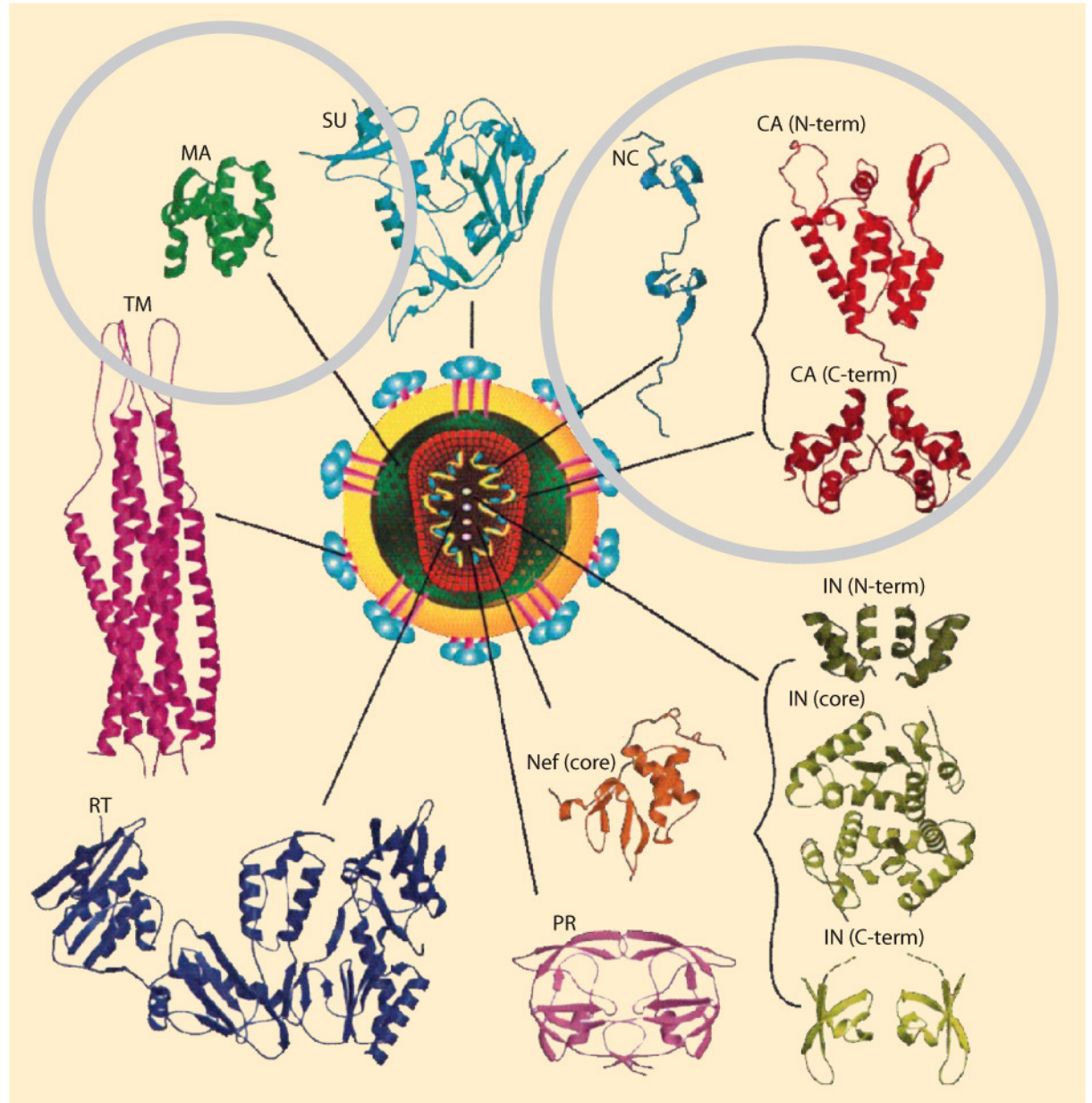
Figure 8.28 Molecular Biology of Assemblies and

tomato bushy stunt virus (TBSV)



# assembly and maturation of human immunodeficiency virus (HIV)

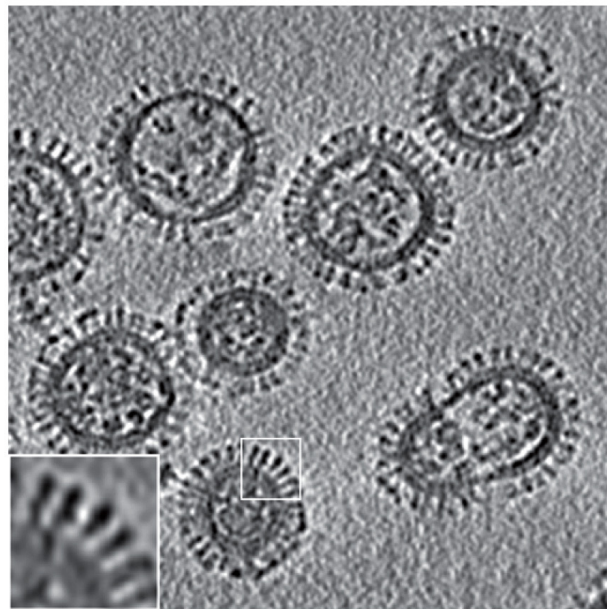




Box 8.1 Figure 8.1.2 Molecular Biology of Assemblies and Machines (© Garland Science 2016)

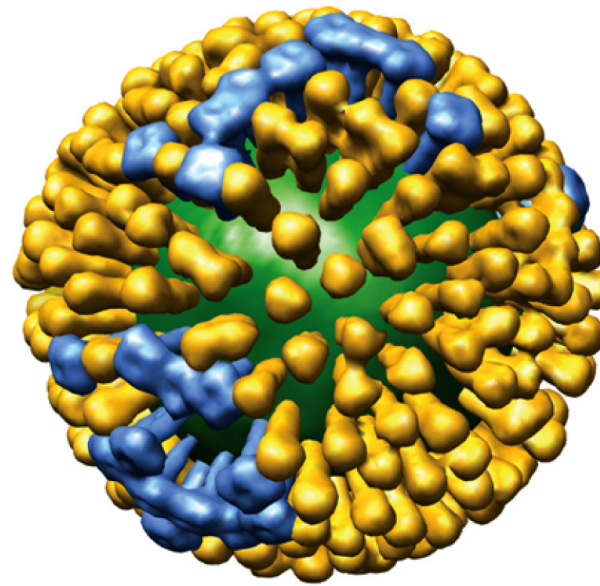


# Influenza virus



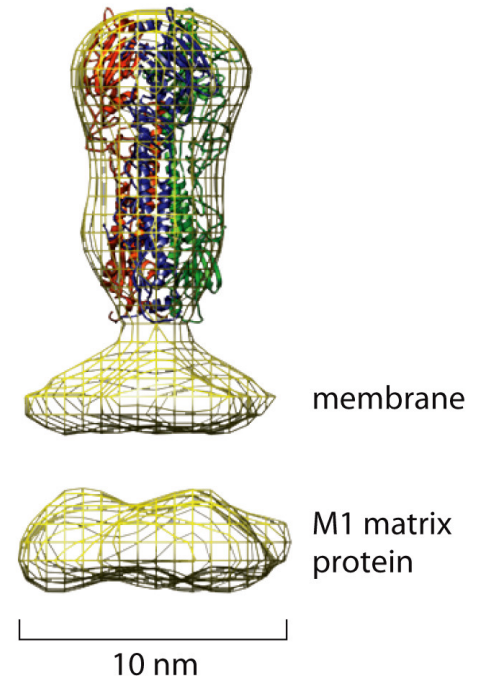
100 nm

Figure 8.60a Molecular Biology of Assemblies and Machines (© Garland Science 2016)



50 nm

Figure 8.60b Molecular Biology of Assemblies and Machines (© Garland Science 2016)

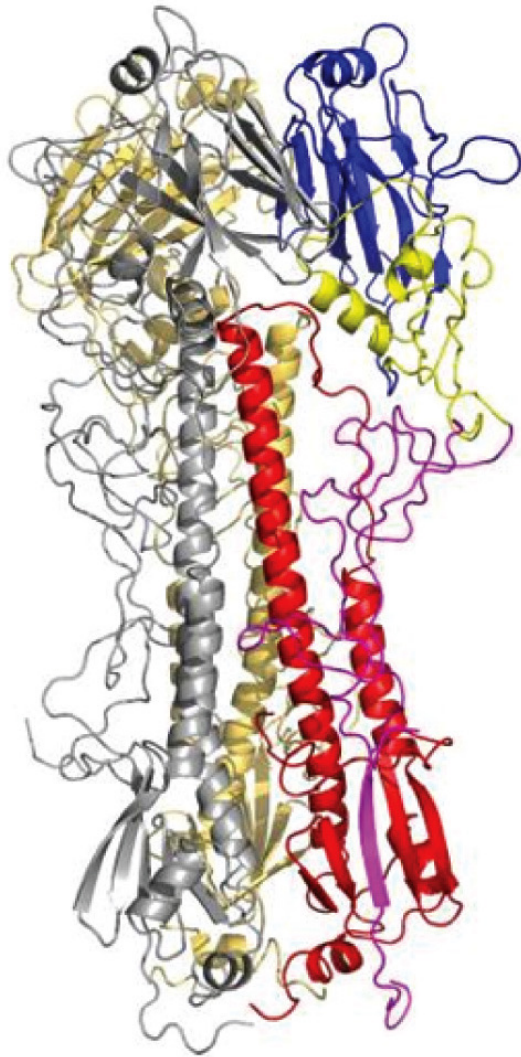


membrane

M1 matrix protein

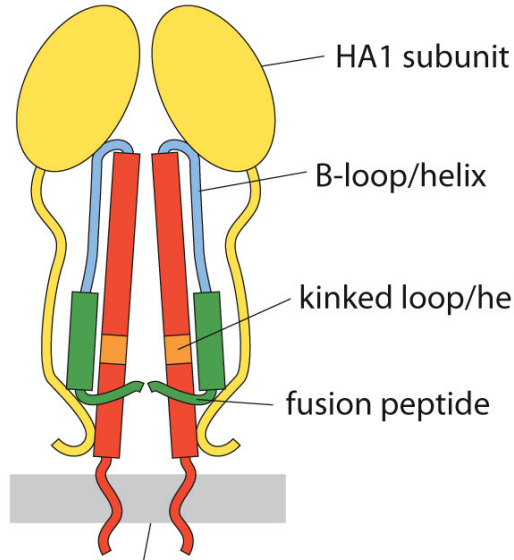
10 nm

Figure 8.60c Molecular Biology of Assemblies and Machines (© Garland Science 2016)

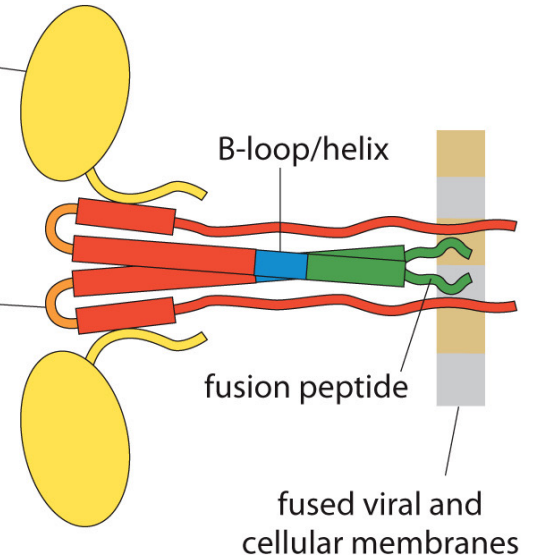


neutral pH/pre-fusion

target membrane



(F) low pH/post-fusion



.60ef Molecular Biology of Assemblies and Machines (© Garland Science 2016)

Figure 8.60d Molecular Biology of Assemblies and Machines (© Garland Science 2016)

# Assembly pathway of influenza virus.

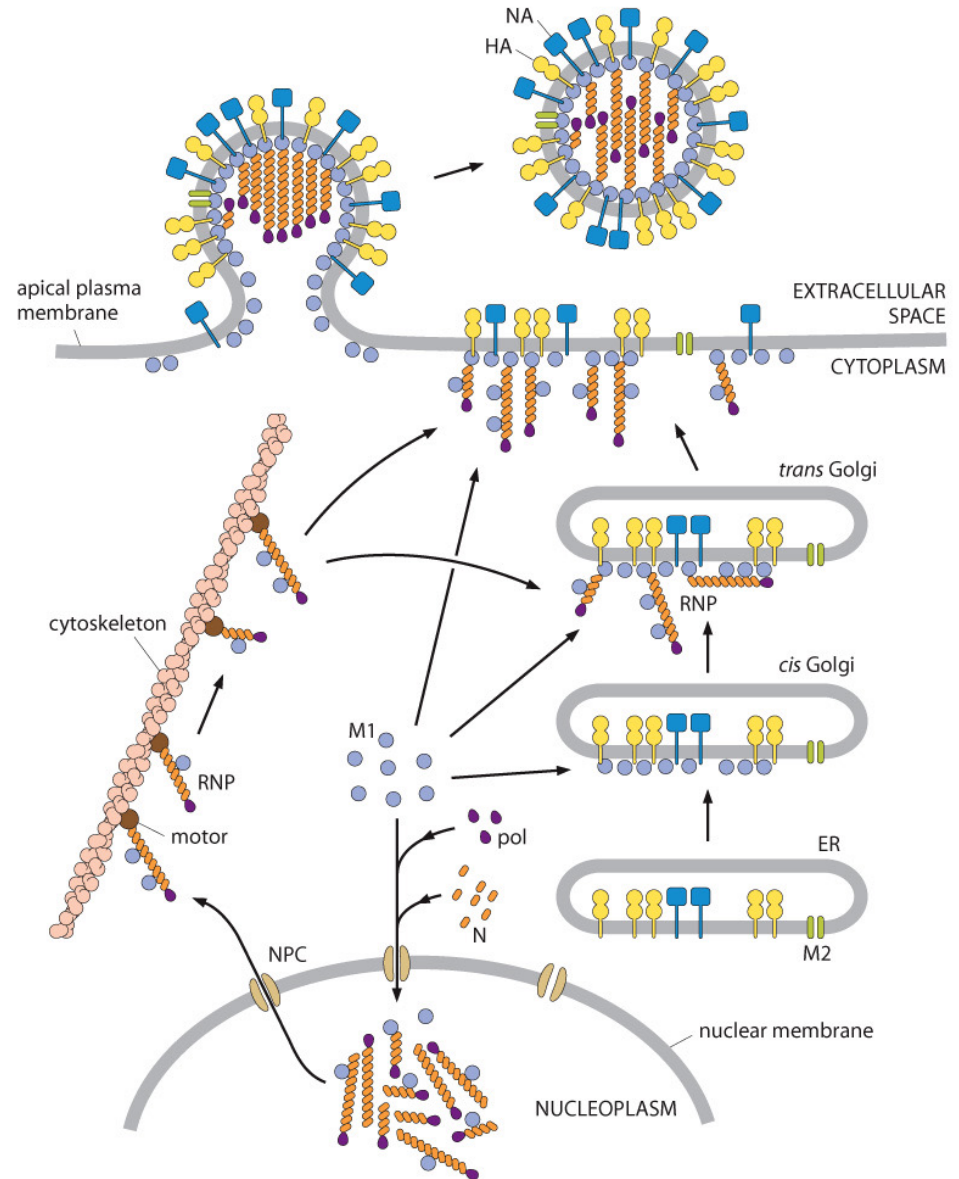
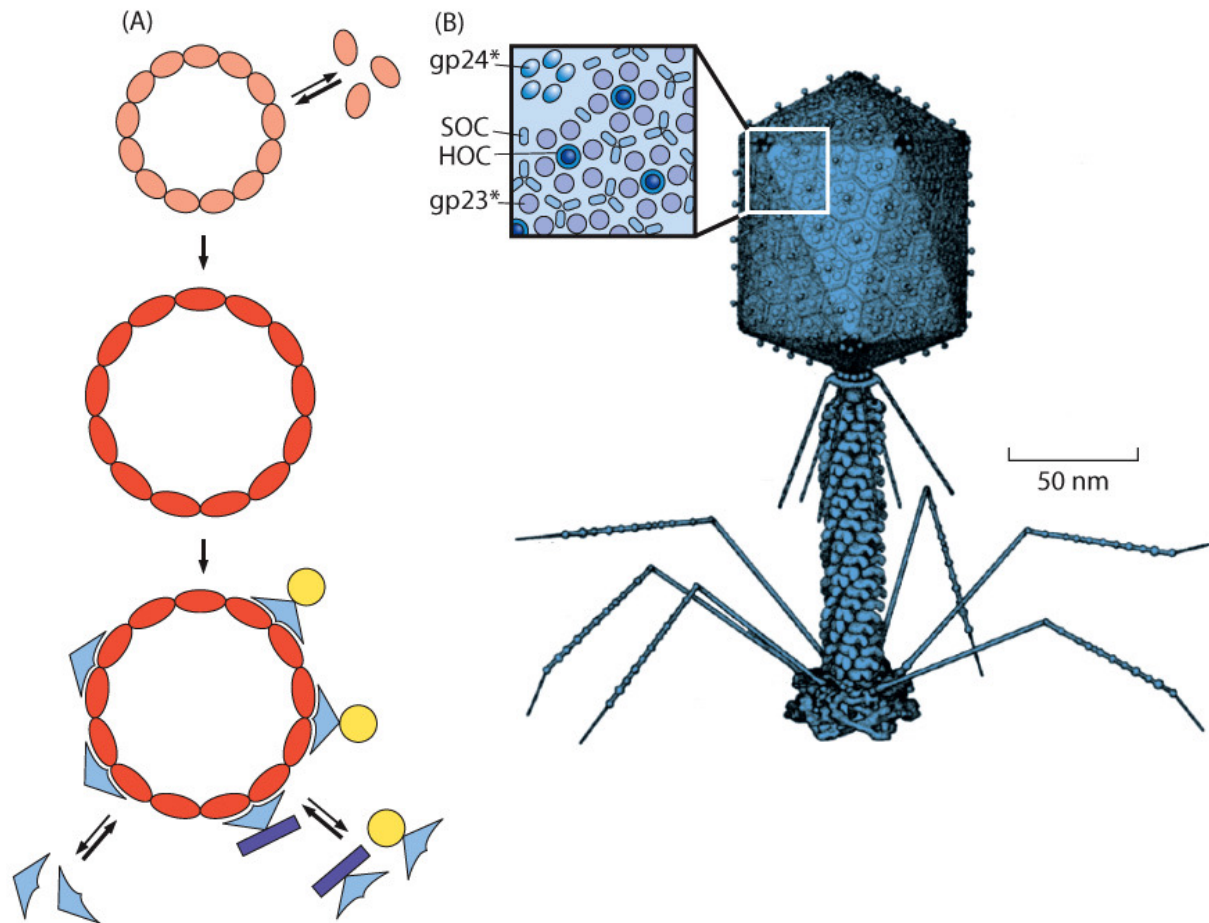


Figure 8.62 Molecular Biology of Assemblies and Machines (© Garland Science 2016)

# Display of proteins on accessory proteins of dsDNA bacteriophages





# Display of proteins on accessory proteins of dsDNA bacteriophages

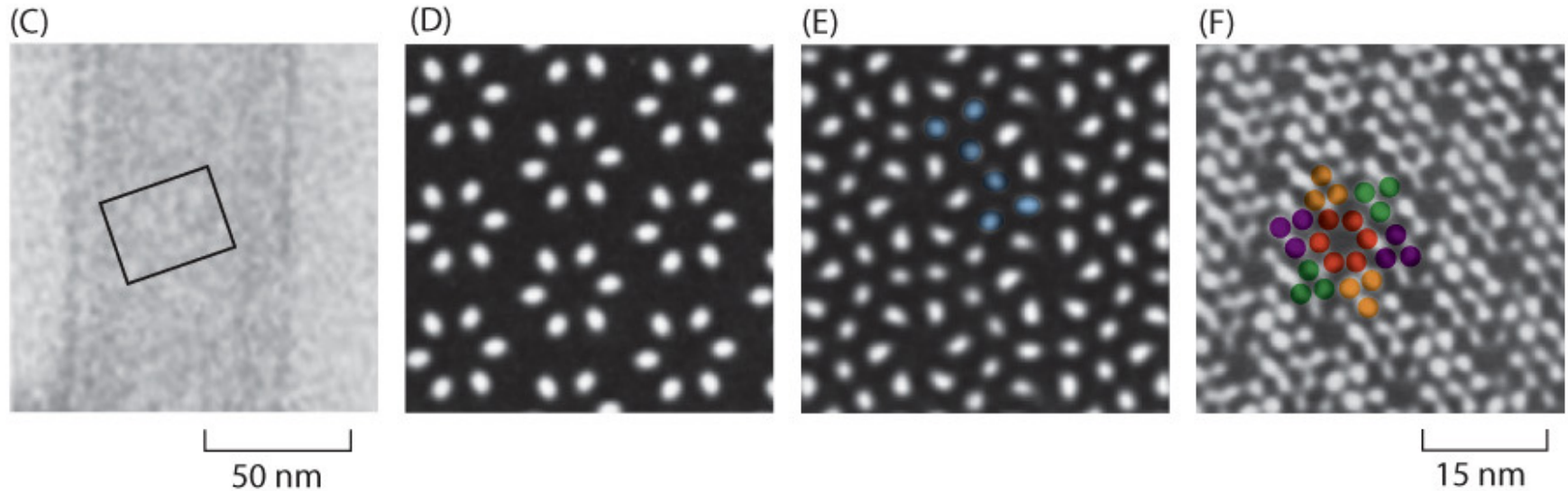


Figure 8.67 Molecular Biology of Assemblies and Machines (© Garland Science 2016)

# Display of an Ig domain

flock house virus

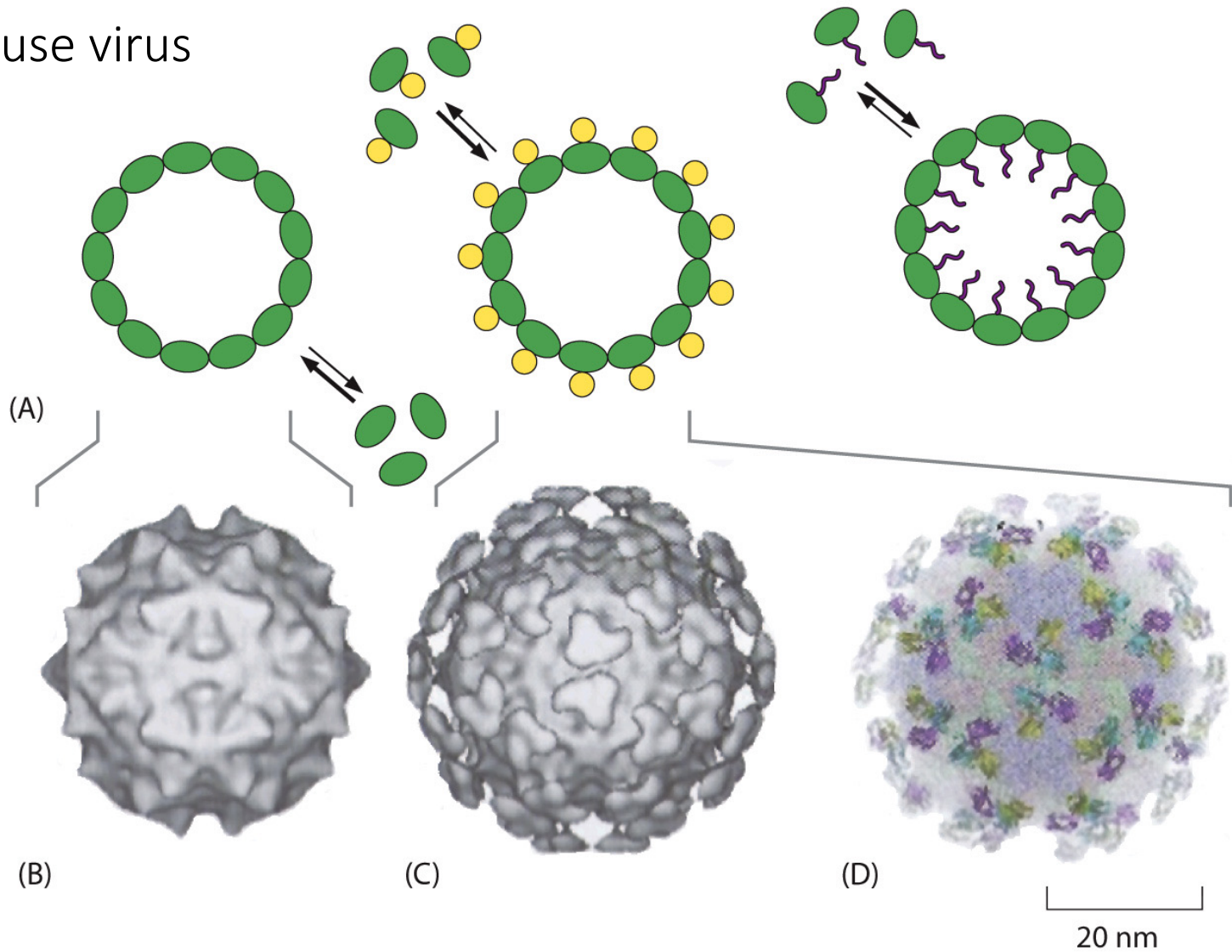


Figure 8.68 Molecular Biology of Assemblies and Machines (© Garland Science 2016)

# Display of green fluorescent protein at the tips of HBV capsid spikes

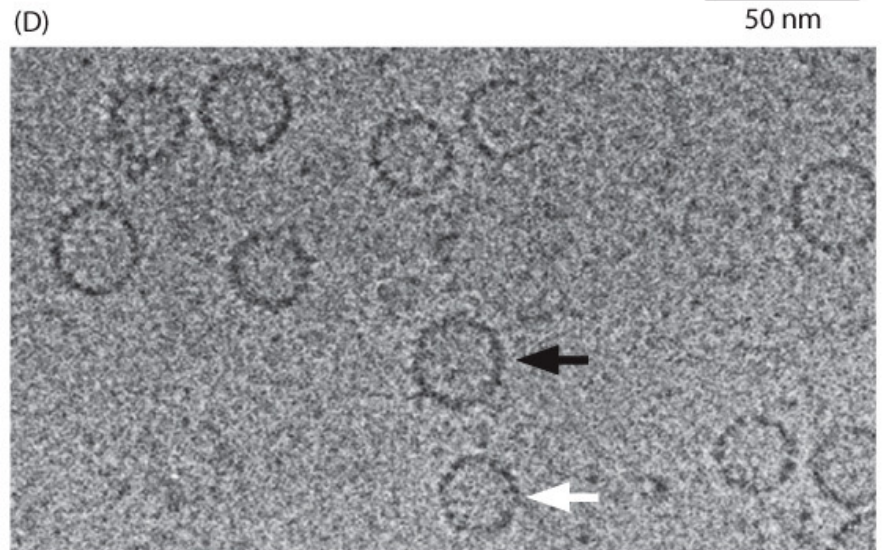
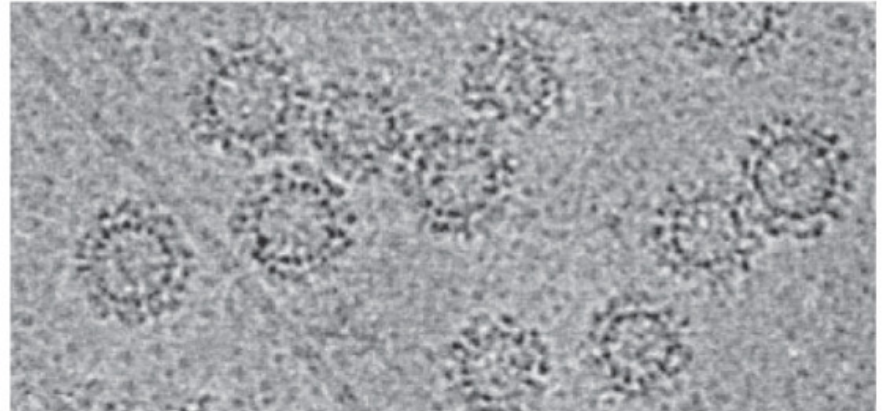
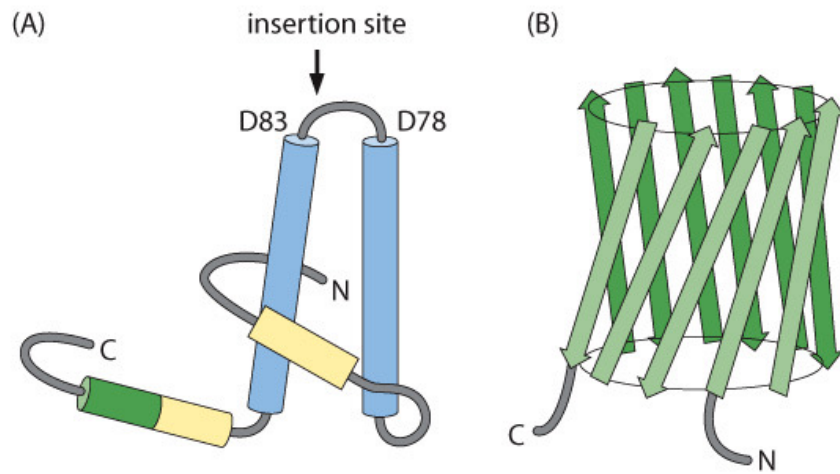


Figure 8.69 Molecular Biology of Assemblies and Machines (© Garland Science 2016)

# Generation of protective vaccines

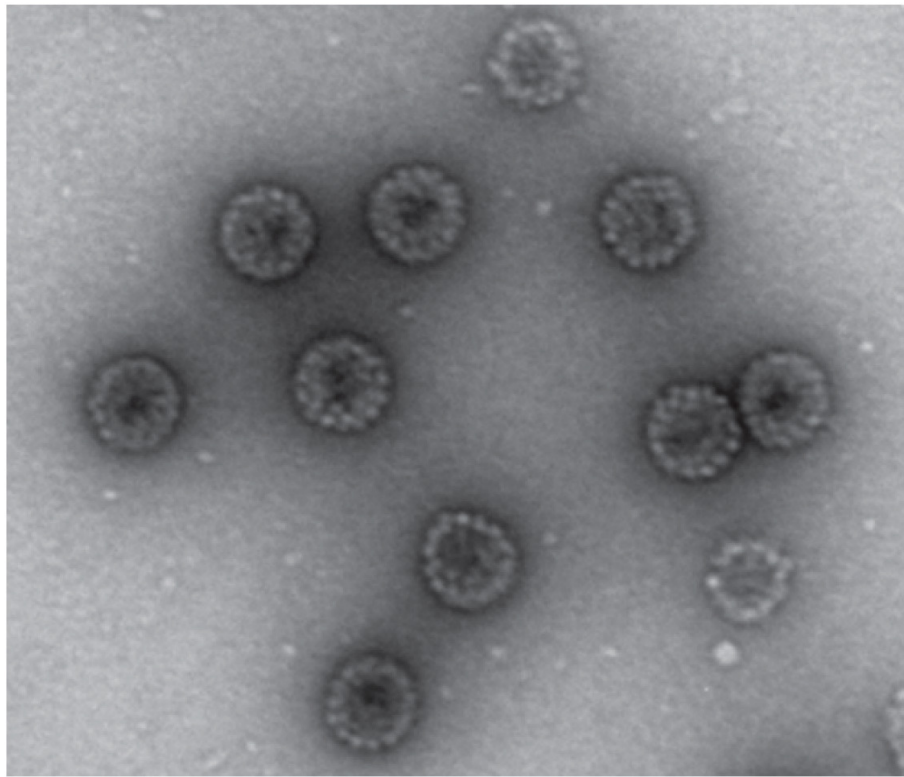


Figure 8.70a Molecular Biology of Assemblies and Machines (© Garland Science 2016)

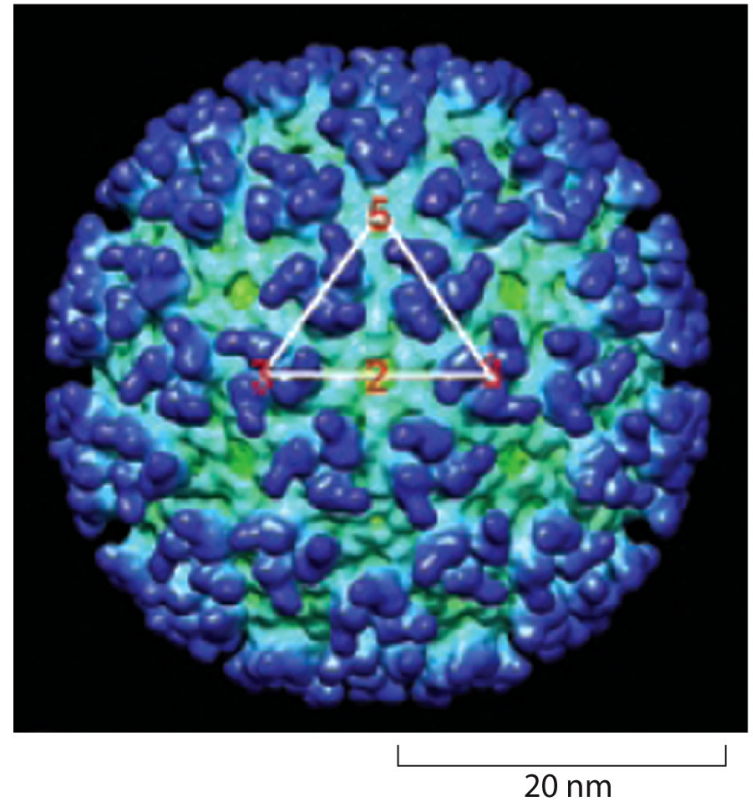


Figure 8.70b Molecular Biology of Assemblies and Machines (© Garland Science 2016)



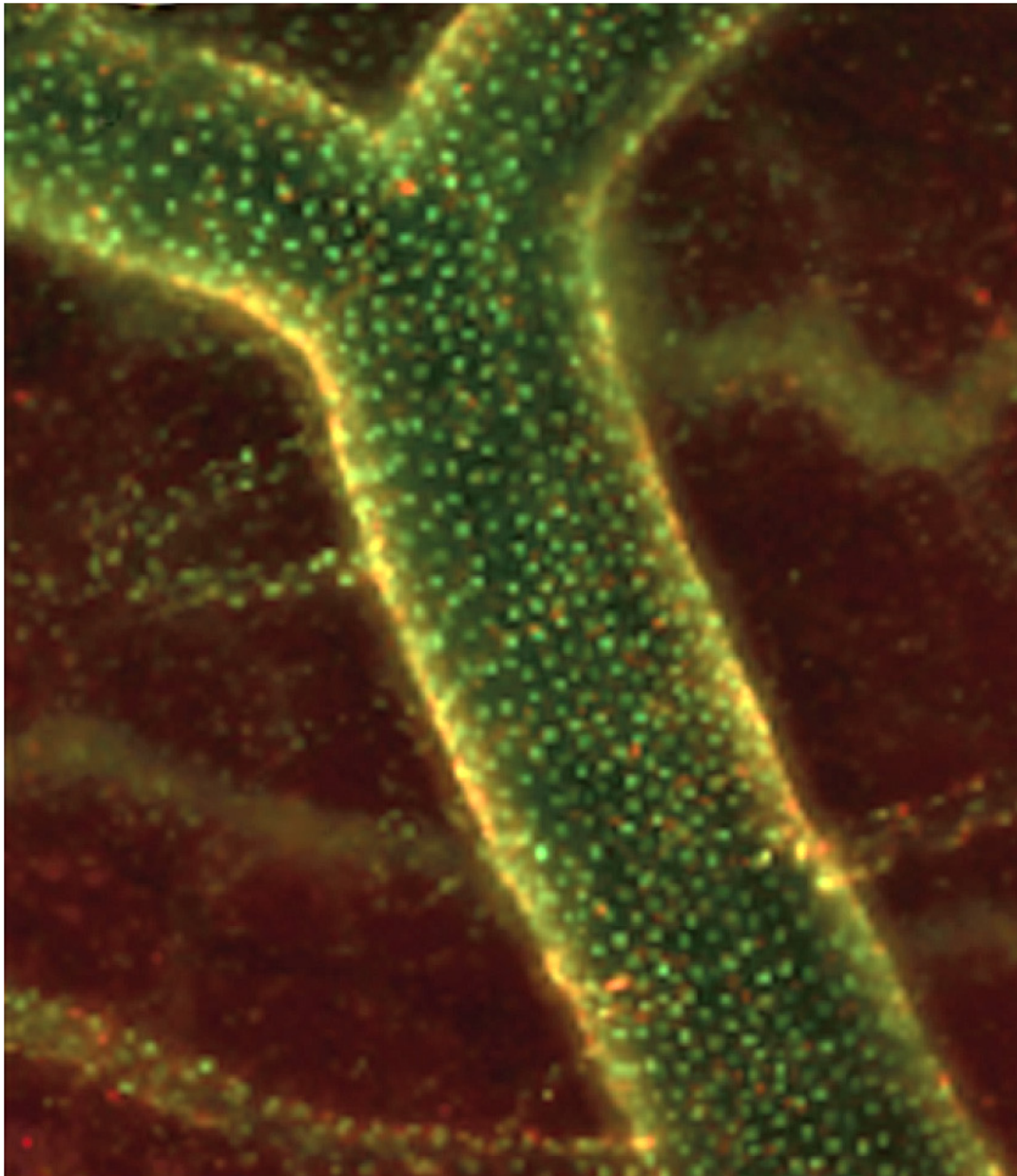


Figure 8.71 Molecular Biology of Assemblies and Machines (© Garland Science 2016)